

The Journal of Parasitology

Vol. 33, No. 6, Section 2 (Supplement)

DECEMBER, 1947

PROGRAM AND ABSTRACTS OF THE TWENTY-SECOND ANNUAL MEETING OF THE AMERICAN SOCIETY OF PARASITOLOGISTS

CHICAGO, ILLINOIS

December 29, 30, and 31, 1947

PROGRAM¹

MONDAY MORNING SESSION, DECEMBER 29, 9:00 AM, CONGRESS HOTEL (FRANCIS I ROOM).

Read

1. Potassium in Relation to Death in Rats Injected with *Trypanosoma equiperdum*. (10 min) GEORGE J. SCHEFF AND JONATHAN S. THATCHER, Chicago Medical School and Ohio State University.

2. Studies on *Bodo foetus* (Protozoa: Mastigophora). (10 min) (Lantern) BANNER BILL MORGAN, University of Wisconsin.

3. Cultivation of *Trichomonas gallinarum* in the Chick Embryo and *in vitro* with Single Strains of Certain Bacteria. (10 min) (Lantern) R. M. CABLE AND EMMA HILLAERT, Department of Biology and Department of Veterinary Science, Purdue University, Lafayette, Indiana.

4. Strains of *Trichomonas gallinae* Varying in Virulence. (7 min) (Lantern) ROBERT M. STABLER, Colorado College.

5. Infection with a Less Virulent Strain of *Trichomonas gallinae* as a Protection against a More Virulent One. (8 min) (Lantern) ROBERT M. STABLER, Colorado College.

6. Unisexual Infections with *Schistosomatium douthitti* (Trematoda). (10 min) ROBERT B. SHORT, University of Michigan.

7. Penetration Glands in Cyclophyllidean Onchospheres. (10 min) (Lantern) W. MALCOLM REID, Monmouth College.

8. Observations on Experimental Dermatitis in Humans Induced by Cercariae of *Trichobilharzia stagnicola* (Talbot, 1936). (15 min) (Lantern) LOUIS OLIVIER, National Institute of Health.

9. Notes on the Life Cycle of *Schistocephalus* sp., a Tapeworm from Gulls. (10 min) (Lantern) LYELL J. THOMAS, University of Illinois.

10. The Sedimentation Rate of Erythrocytes in Malaria Infections and Its Correlation with the Electrophoretic Mobility. (15 min) (Lantern) W. B. REDMOND AND W. S. POLLITZER, Emory University.

¹ An alphabetical author index will be found at the end of the program. Extra copies of this Supplement, and portraits of parasitologists, will be on sale at the meeting.

11. Coccidia of the Mink. (10 min) (Lantern) NORMAN D. LEVINE, University of Illinois.

12. The Effect of Sulfaquinoxaline on the Developmental Stages of *Eimeria tenella*. (10 min) (Lantern) ASHTON C. CUCKLER AND WALTHER H. OTT, Merck Institute for Therapeutic Research, Rahway, New Jersey.

13. Observations on *Leucocytozoon* Infections in Birds Receiving Paludrine, Atebrin, and Sulphamerazine. (10 min) (Lantern) A. MURRAY FALLIS, Ontario Research Foundation, Toronto.

14. A New Ruling for the Ocular Disc of Special Usefulness in the Teaching of Medical Parasitology. (5 min) (Lantern) (Also by demonstration) EDWARD P. OFFUTT, University of Rochester.

MONDAY AFTERNOON SESSION, DECEMBER 29, 2:00 PM, CONGRESS HOTEL (FRANCIS I ROOM).

Read

15. The Metabolism of Mammalian Trypanosomes and their Classification. (10 min) (Lantern) THEODOR VON BRAND AND ELEANOR JOHNSON TOBIE, National Institute of Health.

16. Studies on Experimental Blackhead Infection in Turkeys. (10 min) (Lantern) FRANCIS MCKAY AND NEAL F. MOREHOUSE, Dr. Salsbury's Laboratories, Charles City, Iowa.

17. The Taxonomic Value of the Gnathosome in Certain Laelapine Mites. (10 min) (Lantern) R. W. STRANDTMANN, University of Texas.

18. The Resistance of Certain Adult Ducks to Infection of the Avian Malaria Parasite *Plasmodium lophurae*. (10 min) (Lantern) WILLIAM TRAGER, Rockefeller Institute for Medical Research.

19. Immunity in Avian Malaria in Relation to Severity of Infection. (10 min) (Lantern) WENDELL GINGRICH, University of Texas.

20. Progress Report on the Parasitic Stages of *Chabertia ovina*. (15 min) (Lantern) (Also by demonstration) W. L. THRELKELD, Virginia Agricultural Experiment Station, Blacksburg, Virginia.

21. What is a Virgula in Virgulate Cercariae? (15 min) (Motion picture) (Also by demonstration) FRANCIS KRUIDENIER, University of Michigan.

22. Quantitative Hookworm Diagnosis by Direct Smear. (15 min) (Lantern) (Also by demonstration) PAUL C. BEAVER, Tulane University.

23. A Method of Testing the Effect of Substances upon the Activity of the Larger Nematodes. (10 min) (Lantern) K. B. KERR AND J. W. CAVETT, Dr. Salsbury's Laboratories, Charles City, Iowa.

24. Immunity in Chickens to *Ascaridia galli*. (15 min) (Lantern) ELVIO H. SADUN, Johns Hopkins University.

25. Factors in the Resistance of White Rats to Infection with the Cotton Rat Filariids. (15 min) (Lantern) J. ALLEN SCOTT, University of Texas.

26. Comparative Filaricidal Activity of Trivalent Arsenic and Antimony. (15 min) (Lantern) G. F. OTTO AND T. H. MAREN, Johns Hopkins University.

TUESDAY MORNING SESSION, DECEMBER 30, 9:00 AM, CONGRESS HOTEL (FRANCIS I ROOM).

Read

27. Resumption of the Reproductive Phase of *Trypanosoma lewisi* in Recovered and Near-Recovered Rats Following both Reinoculation with the Parasite and Treatment with Sodium Salicylate. (15 min) (Lantern) ELERY R. BECKER AND MICHAEL G. LYSENKO, Iowa State College.

28. An Effective New Coccidiostatic. (15 min) (Opaque projection) PAUL D. HARWOOD, DOROTHY I. STUNZ, AND ROBERT WOLFGANG, Dr. Hess and Clark, Inc., Ashland, Ohio.

29. A Method for Screening Anti-Malarial Drugs in the Mosquito Host. (15 min) (Lantern) LEVON A. TERZIAN, Naval Medical Research Institute.

30. Observations on Oribatid Mites, Vectors of *Moniezia expansa* on Pastures, with a Report of Several New Vectors from the United States. (15 min) (Lantern) (Also by demonstration) K. C. KATES AND C. E. RUNKEL, U. S. Bureau of Animal Industry.

31. The Relationship between Trombiculid and Trombidid Mites. (15 min) (Lantern) G. W. WHARTON, Duke University.

32. Extended Persistence of Oöcysts of *Plasmodium relictum* in Culture. (10 min) (Lantern) GORDON H. BALL, University of California at Los Angeles.

33. The Infectivity of Native Malarias in South Carolina to *Anopheles quadrimaculatus*. (10 min) (Lantern) MARTIN D. YOUNG, NEWTON F. HARDMAN, ROBERT W. BURGESS, WILLIAM C. FROHNE, AND CURTIS W. SABROSKY, U. S. Public Health Service.

TUESDAY MORNING SESSION, DECEMBER 30, 11:00 AM, CONGRESS HOTEL (FRANCIS I ROOM).

Presidential Address

34. Expanding Horizons in the Recognition of a Phylum. HARLEY J. VAN CLEEVE, University of Illinois.

TUESDAY, DECEMBER 30.

12:30 PM, PARASITOLOGISTS LUNCHEON.

1:30 PM, ANNUAL BUSINESS MEETING.

TUESDAY AFTERNOON SESSION, DECEMBER 30, 3:00 PM, CONGRESS HOTEL (FRANCIS I ROOM).

By Demonstration

14. A New Ruling for the Ocular Disc of Special Usefulness in the Teaching of Medical Parasitology. (Also read) EDWARD P. OFFUTT, University of Rochester.

20. Progress Report on the Parasitic Stages of *Chabertia ovina*. (Also read) W. L. THRELKELD, Virginia Agricultural Experiment Station, Blacksburg, Virginia.

21. What Is a Virgula in Virgulate Cercariae? (Also read) FRANCIS KRUDENIER, University of Michigan.

22. Quantitative Hookworm Diagnosis by Direct Smear. (Also read) PAUL C. BEAVER, Tulane University.

30. Observations in Oribatid Mites, Vectors of *Moniezia expansa* on Pastures, with a Report of Several New Vectors from the United States. (Also read) K. C. KATES AND C. E. RUNKEL, U. S. Bureau of Animal Industry.

35. The Trematode Genus *Tamerlania* in Resident Birds of the United States. ELON E. BYRD, University of Georgia.

36. Mucin in Developing Digenetic Trematodes. FRANCIS J. KRUIDENIER, University of Michigan.

37. Some Effects of Acanthocephala on the Large-Mouthed Black Bass. CARL E. VENARD AND JOHN H. WARFEL, Ohio State University and Western Reserve University.

38. A Method for Relaxing and Fixing Large Cestodes. ROBERT L. ROUDA-BUSH, Ward's Natural Science Establishment, Inc.

53. The Phase Microscope in the Examination of *Endamoeba*. (Also read) ERNEST HARTMAN AND GEORGE F. SCHEFF, Chicago Medical School.

WEDNESDAY MORNING SESSION, DECEMBER 31, 9:00 AM, CONGRESS HOTEL (FRANCIS I ROOM).

Read

39. Mammalian Blood Flukes of Celebes. (10 min) (Lantern) ERNEST CARROLL FAUST, Tulane University, AND C. BONNE, Universiteit van Indonesie, Batavia, Java.

40. Germinal Masses in the Rediae of the Trematode Order Fasciolatoidea Szidat, 1936. (15 min) (Lantern) W. W. CORT, D. J. AMEEL, AND ANNE VAN DER WOUDE, Johns Hopkins University, Kansas State College, and University of Michigan.

41. Germinal Material in Mother Sporocysts and Rediae of *Halipegus eccentricus* Thomas, 1939. (10 min) (Lantern) D. J. AMEEL, W. W. CORT, AND ANNE VAN DER WOUDE, Kansas State College, Johns Hopkins University, and University of Michigan.

42. *Porocephalus crotali* Humbolt (Pentastomida) in Oklahoma. (8 min) (Lantern) J. TEAGUE SELF, University of Oklahoma.

43. Additional Infection Experiments with the Hookworm, *Bunostomum phlebotomum*, in the Calf. (15 min) ROY L. MAYHEW, Louisiana State University.

44. On Certain Pseudophyllidean Cestodes from Alaskan Pinnipeds. (12 min) HORACE W. STUNKARD, New York University.

45. Nature of Duodenal Nematode Growth-Inhibiting Factor. (10 min) (Lantern) LYMAN P. FRICK AND J. E. ACKERT, Kansas State College.

46. Further Studies on Hydrogen-Ion Concentration as a Factor in Age Resistance to the Fowl Ascarid. (10 min) (Lantern) B. B. RIEDEL AND J. E. ACKERT, Kansas State College.

47. Age of *Ascaridia* Egg Cultures a Factor in Worm Growth. (5 min) (Lantern) J. E. ACKERT, RHODA W. COOPER, AND L. W. DEWHIRST, Kansas State College.

48. The Status of the Sodium Fluoride Treatment for the Removal of Large Roundworms from Swine. (15 min) (Lantern) A. O. FOSTER, F. D. ENZIE, R. T. HABERMANN, AND R. W. ALLEN, U. S. Bureau of Animal Industry.

49. Screening Large Numbers of New Chemical Compounds for Anthelmintic Activity Using Infections of *Nippostrongylus muris* in Mice. (15 min) (Lantern) STERLING BRACKETT AND ALEXANDER BLIZNICK, American Cyanamid Company, Stamford, Connecticut.

50. Further Studies on the Maintenance of Cultures of *Endamoeba histolytica* without Viable Bacteria. (10 min) (Lantern) LEON JACOBS, National Institute of Health.

51. Studies on the Growth Requirements of Metacystic *Endamoeba histolytica*. (10 min) (Lantern) CHARLES W. REES AND LUCY V. REARDON, National Institute of Health.

52. Culture Experiments on Intestinal Flagellates. IV. Selected Longevity Records to September 15, 1947. (15 min) (Lantern) D. H. WENRICH, University of Pennsylvania.

53. The Phase Microscope in the Examination of *Endamoeba*. (7 min) (Also by demonstration) ERNEST HARTMAN AND GEORGE J. SCHEFF, Chicago Medical School.

54. The Fate of Various Species of Trypanosomes in *Triatoma*. (10 min) (Lantern) A. PACKCHANIAN, University of Texas.

55. Studies of Sheep Parasites. VIII. Overwintering of Nematode Larvae (10 min) (Lantern) PHILIP A. HAWKINS AND MOACYR G. DE FREITAS.

By Title

56. The Effect of Aralen on *Giardia lamblia* Infections in Children. J. C. SWARTZWELDER AND T. C. PAPERMASTER, Louisiana State University.

57. Bite of *Amblyomma americanum* Associated with Possible Tick Paralysis. J. C. SWARTZWELDER AND J. H. SEABURY, Louisiana State University.

58. Report of Ten Cases of *Balantidium coli* Infection. J. C. SWARTZWELDER, Louisiana State University.

59. The Life History and Bionomics of *Heterakis bonasae*, a Cecal Nematode of Bobwhite Quail, *Colinus virginianus* and *C. virginianus texanus*. J. W. WARD, University of Mississippi.

60. Further Taxonomic Studies on Internal Parasites of Horses and Mules. J. W. WARD, University of Mississippi.

61. Studies on Parasites and Food Habits of Foxes. J. W. WARD, University of Mississippi.

62. Longevity and Rates of Infectivity of the Free-Swimming Miracidia of *Schistosoma mansoni*. JOSE F. MALDONADO AND J. ACOSTA-MATIENZO, School of Tropical Medicine, San Juan, Puerto Rico.

63. An Investigation of Some Possible Sources of Trichina Infection in a Central Illinois Community. WAYNE W. WANTLAND AND PAUL MARTIN, Illinois Wesleyan University.

64. The Localization of Adult *Trichinella spiralis* in the Intestinal Tract of Young and Old Mice. JOHN E. LARSH, JR. AND JAMES R. HENDRICKS, University of North Carolina.

65. The Effect of Thyroid and Thiouracil on the Natural Resistance of Mice to Infection with *Hymenolepis*. JOHN E. LARSH, JR., University of North Carolina.

66. The White Rat and the Guinea Pig as Hosts for the Larvae of the Brown Dog Tick, *Rhipicephalus sanguineus*. GEORGE W. LUTTERMOSER, University of Pennsylvania.

67. Susceptibility and Immunity of Marine Fishes to *Benedenia* (= *Epibdella*) *melleni* (MacCallum), a Monogenetic Trematode. III. Natural Hosts in the West Indies. ROSS F. NIGRELLI, New York Aquarium, New York Zoological Society.

68. Parasites of the Ranidae (Amphibia). I. A. C. WALTON, Knox College.
69. Parasites of the Ranidae (Amphibia). II. A. C. WALTON, Knox College.
70. Parasites of the Ranidae (Amphibia). III. A. C. WALTON, Knox College.

WEDNESDAY AFTERNOON SESSION, DECEMBER 31, 2:00 PM, CONGRESS HOTEL
(FRANCIS I ROOM).

Symposium on Exoerythrocytic Forms of Malarial Parasites

71. Introduction. CLAY G. HUFF, Naval Medical Research Institute.
72. A Search for the Pre-erythrocytic Stages of *Plasmodium vivax* and of *P. cynomolgi*. CLAY G. HUFF AND FREDERICK COULSTON, Naval Medical Research Institute, and E. I. du Pont de Nemours and Company.
73. The Chemotherapy of Malaria in Relation to Our Knowledge of Exoerythrocytic Forms. G. ROBERT COATNEY AND W. CLARK COOPER, National Institute of Health.
74. The Chemotherapy and Immunology of Pre-erythrocytic Stages in Avian Malaria. FREDERICK COULSTON AND CLAY G. HUFF, E. I. du Pont de Nemours and Company and Naval Medical Research Institute.
75. RICHARD JANVIER PORTER, University of Michigan. (Invited Paper; No Abstract Received.)
76. Response of Exo-erythrocytic Forms to Alterations in the Life Cycle of *Plasmodium gallinaceum*. VICTOR H. HAAS, AIMEE WILCOX, R. L. LAIRD, FRANCES MOORE EWING, AND NELL COLEMAN, National Institute of Health and University of Tennessee.

AUTHOR INDEX

Showing program number, which is also abstract number of each paper.

<i>Author</i>	<i>Program and Abstract Number</i>	<i>Author</i>	<i>Program and Abstract Number</i>
Ackert, J. E.	45, 46, 47	Martin, P.	63
Acosta-Matienzo, J.	62	Mayhew, R. L.	43
Allen, R. W.	48	McKay, F.	16
Ameel, D. J.	40, 41	Morehouse, N. F.	16
Ball, G. H.	32	Morgan, B. B.	2
Beaver, P. C.	22	Nigrelli, R. F.	67
Becker, E. R.	27	Offutt, E. P.	14
Bonne, C.	39	Olivier, L.	8
Bliznick, A.	49	Ott, W. H.	12
Brackett, S.	49	Otto, G. F.	26
v. Brand, T.	15	Packchianian, A.	54
Burgess, R. W.	33	Papermaster, T. C.	56
Byrd, E. E.	35	Pollitzer, W. S.	10
Cable, R. M.	3	Porter, R. J.	75
Cavett, J. W.	23	Reardon, L.	51
Coatney, G. R.	73	Redmond, W. B.	10
Coleman, N.	76	Rees, C. W.	51
Cooper, W.	47	Reid, W. M.	7
Cooper, W. C.	73	Riedel, B. B.	46
Cort, W. W.	40, 41	Roudabush, R. L.	38
Coulston, F.	72, 74	Runkel, C. E.	30
Cuckler, A. C.	12	Sabrosky, C. W.	33
Dewhirst, L. W.	47	Sadun, E. H.	24
Enzie, F. D.	48	Scheff, G. J.	1, 53
Ewing, F. M.	76	Scott, J. A.	25
Fallis, A. M.	13	Seabury, J. H.	57
Faust, E. C.	39	Self, J. T.	42
Foster, A. O.	48	Short, R. B.	6
de Freitas, M. G.	55	Stabler, R. M.	4, 5
Frick, L. P.	45	Strandtmann, R. W.	17
Frohne, W. C.	33	Stunkard, H. W.	44
Gingrich, W.	19	Stunz, D. I.	28
Haas, V. H.	76	Swartzwelder, J. C.	56, 57, 58
Habermann, R. T.	48	Terzian, L. A.	29
Hardman, N. F.	33	Thatcher, J. S.	1
Hartman, E.	53	Thomas, L. J.	9
Harwood, P. D.	28	Threlkeld, W. L.	20
Hawkins, P. A.	55	Tobie, E. J.	15
Hendricks, J. R.	64	Trager, W.	18
Hillaert, E.	3	Van Cleave, H. J.	34
Huff, C. G.	71, 72, 74	Van der Woude, A.	40, 41
Jacobs, L.	50	Venard, C. E.	37
Kates, K. C.	30	Walton, A. C.	68, 69, 70
Kerr, K. B.	23	Wantland, W. W.	63
Kruidenier, F. J.	21, 36	Ward, J. W.	59, 60, 61
Laird, R. L.	76	Warfel, J. H.	37
Larsh, J. E.	64, 65	Wenrich, D. H.	52
Levine, N. D.	11	Wharton, G. W.	31
Luttermoser, G. W.	66	Wilcox, A.	76
Lysenko, M. G.	27	Wolfgang, R.	28
Maldonado, J. F.	62	Young, M. D.	33
Maren, T.	26		

ABSTRACTS

1. *Potassium in Relation to Death in Rats Infected with Trypanosoma equiperdum*. GEORGE J. SCHEFF AND JONATHAN S. THATCHER, Chicago Medical School and Ohio State University.

Potassium tolerant and non-tolerant rats were studied with respect to the progress of the parasite count and survival time following infection with *Trypanosoma equiperdum*. From the observation that in both groups of animals the course of the infection was practically the same, it appears that potassium is not the causative agent of death in this disease as claimed by some recent investigators.

2. *Studies on Bodo foetus (Protozoa: Mastigophora)*. BANNER BILL MORGAN, University of Wisconsin.

Bodo foetus, a flagellate originally isolated and described by Koffman (1937) from aborted bovine fetuses in Sweden, has been found to be the most common contaminant in samples collected for the diagnosis of bovine trichomoniasis. Attempts to isolate *B. foetus* in bacteria-free cultures have been negative. This flagellate has been maintained in the laboratory for four years in association with a single species of bacteria (*Proteus* sp.) by weekly transfers. *Bodo foetus* grows best at 30°C in egg slants overlaid with Ringer's solution or in a medium composed of peptone, ammonium nitrate, sodium acetate, potassium phosphate, magnesium sulfate and distilled water. The optimum pH is 6.6. *Bodo foetus* will die within 24 to 36 hours at 37°C. Inoculations (intravenous, intramuscular, intraperitoneal and subcutaneous) of small laboratory animals were routinely negative. Vaginal inoculations into cows were also negative. It was concluded that *B. foetus* is a non-pathogenic, coprozoic flagellate.

3. *Cultivation of Trichomonas gallinarum in the Chick Embryo and in vitro with Single Strains of Certain Bacteria*. R. M. CABLE AND EMMA HILLAERT, Purdue University.

Trichomonas gallinarum was grown in the chorio-allantoic fluid of living chick embryos injected with the cecal contents of infected turkeys and protected against massive bacterial infection by the addition of 5000 units of penicillin/cc of inoculum. Flagellates persisted in the chorio-allantoic fluid until hatching and were recovered from the intestine of the newly hatched chicks. *T. gallinarum* has been passed successively through embryos 23 times at one- to two-week intervals and apparently can be maintained in this manner indefinitely, although the chorio-allantoic fluid is not a favorable medium; only occasionally did the flagellates become numerous in it. By successive passage and selection of embryos in which the flagellates grew best, they were obtained bacteria-free and used in two series of cultivation experiments. In these, small numbers of organisms were added to liver-infusion-agar slants overlaid with turkey serum-saline, with and without single strains of certain bacteria. The flagellates failed to multiply in the medium alone or in medium plus *Escherichia coli* or *Aerobacter aerogenes*, whereas they became abundant in the presence of *Brucella suis*, *Pseudomonas fluorescens*, or *Bacillus subtilis*. They attained the highest counts and persisted longest, as a rule, in cultures with *B. suis*.

4. *Strains of Trichomonas gallinae Varying in Virulence*. ROBERT M. STABLER, Colorado College.

The varying reactions of similarly housed pigeons, from the asymptomatic carrier to the bird dying riddled with canker lesions, has led to speculation as to whether or not there exists more than one strain of *T. gallinae* with respect to virulence. To test this idea, the writer used the flagellates from the mouths of pigeons from 5 widely separated and unrelated sources. These 5 strains were put successively into the mouths of 5 series each of 5 exactly comparable pigeons. They were 6, 5½, 22, 28, and 36 weeks of age, respectively. The strains were numbered as I-V; the 5 series of birds as 1-5; and the extent of lesion recorded as 0=no gross pathology, +=very slight oral caseation which healed in 2-4 days, ++=moderate caseation lasting about 10 days, +++=severe caseation remitting just short of death, and ++++=extreme caseation terminating in death. The results were as follows: strain I gave ++, +, +, +, +, and +++ respectively for the five runs; strain II gave 0, 0, 0, 0, and 0 lesions; strain III gave 0, +, 0, +, and + respectively; strain IV showed +, ++, +, +, and + lesions; while strain V gave +, ++, ++, ++, and ++. It appears, then that these strains of *T. gallinae* varied from one which produced no visible pathology (II), to one that actually killed 4 of the 5 birds into which it was put (I).

5. *Infection with a Less Virulent Strain of Trichomonas gallinae as a Protection Against a More Virulent One*. ROBERT M. STABLER, Colorado College.

Having established the variation in virulence among strains of *T. gallinae* it seemed worth determining whether or not infection with a milder strain would protect against the effects of a more virulent one. Accordingly, 8 pigeons 7 wks. old were inoculated *per os* with a strain (V) which always caused severe canker, and some deaths. In 3 days all showed marked lesions,

and 2 died on the 8th and 13th days, respectively. The remaining 6 recovered. They were each given a more severe strain (I) on the 54th day after the initial inoculation. This strain (I) had just killed 12 of 13 birds into which it had been put. None of these 6 birds developed the slightest gross lesion, and on autopsy at the 83rd day all were free of active pathology. Eight more birds, 7 at 8 wks. and one at 2 yrs. of age, were given a strain (III) that had never produced more than an occasional, short-lived caseation. Only one (at 13 days) showed a small, quickly-disappearing canker spot. At 27 days these were given the virulent strain (I). One bird showed a lesion at 5 days, was quite cankerous at 8, and had recovered by 13 days. A second showed a small, short-lived lesion at 13 days. No bird died. On autopsy 21 days after receiving strain I, the bird with the severe pathology at 8 days had considerable liver scarring; the other 7 showed nothing. It would seem that some protection against a very virulent strain of *T. gallinae* was acquired with recovery from an attack by a less virulent strain.

6. *Unisexual Infections with Schistosomium douthitti (Trematoda)*. ROBERT B. SHORT, University of Michigan.

Three white rats and eight laboratory-reared mice (*Peromyscus maniculatus*) were exposed to cercariae of *Schistosomium douthitti* from two wild snails (*Lymnea palustris*)—one naturally, the other experimentally infected. At autopsy, 23–81 days after the first exposure, all hosts were positive and only female worms were recovered. These were sexually mature, for eggs were in uterus and in host tissues. In seven infections of 32 days duration or longer, some eggs in host tissues contained fully developed, living miracidia; a few hatched under coverglass pressure.

Later, 10 laboratory-reared *L. palustris*, each infected by a single miracidium derived from bisexual infections, were sources of cercariae for infection of laboratory-reared *P. maniculatus*. All mice herein considered harbored adult worms of one sex only. Only males developed from cercariae coming from five snails and females from the other five. Sexual maturity of males and females was attained in the absence of the other sex.

In this series, 22 mice were parasitized by females only; these contained eggs in uterus, and fully developed, active miracidia were found in eggs in the tissues of 16 mice infected for 29 days or more.

No spermatozoa were found in the seminal receptacles of 20 stained females from these unisexual infections (14 mounted whole, six serially sectioned) nor in 100 eggs in the six worms sectioned.

This appears to be the first report on the Schistosomatidae involving the full development of female worms and their ova in the absence of males.

7. *Penetration Glands in Cyclophyllidean Onchospheres*. W. MALCOLM REID, Monmouth College.

A pair of unicellular glands has been demonstrated in the onchospheres of the cestodes, *Railletina cesticillus* (Molin) and *Choanotaenia fundibulum* (Blösch) from chickens, *Moniezia expansa* (Rudolphi) from sheep, and *Hymenolepis* sp. from herring gulls. These glands stretch from the posterior end of the embryo forward to pores at the anterior tip where the granular secretion sometimes exudes during larval movements. The uninucleate glands are connected by a narrow isthmus near the posterior end.

Functionally these glands appear to be correlated with the penetration process. The presence of this secretion near the area of hook activity suggests that it assists either by erosion of the cells in the area of invasion or by means of its adhesive properties in helping the hooks to obtain a better purchase on the host tissues. Although similar penetration glands have been recognized in trematode cercariae and miracidia, they have not been generally recognized in cestode larvae.

8. *Observations on Experimental Dermatitis in Humans Induced by Cercariae of Trichobilharzia stagnicolae* (Talbot, 1936). LOUIS OLIVIER, National Institute of Health.

Twenty-two individuals were exposed to cercariae of *Trichobilharzia stagnicolae*, one of the known dermatitis-producing cercariae. From 10 to 30 cercariae were applied to the skin of the forearm within a circular area approximately 18 millimeters in diameter. Evidence of penetration was obtained in each case. In all but 3 cases a distinct pricking sensation was experienced as the cercariae penetrated. In all cases macules were observed in the exposed area within a few hours after the cercariae were applied. The macules usually disappeared within a few hours after their appearance. Four of the 22 individuals had a history of schistosome dermatitis contracted naturally. In each of these 4 cases typical schistosome dermatitis developed after the cercariae were applied in this test. The other 18 individuals had no history of exposure to cercariae of *T. stagnicolae* or other dermatitis-producing cercariae. In none of these 18 cases did typical schistosome dermatitis develop. Three of the 18 had no lesions following the disappearance of the macules. Papules developed in the remaining 15 cases but these papules were

strikingly different from those described for typical schistosome dermatitis. They differed in that they were smaller, they appeared later, and there was no edema or diffuse erythema associated with them. Moreover, in only 2 of the cases was there any itching and in those it was mild and transitory. These results suggest that the clinical entity known as schistosome dermatitis does not develop when an individual is exposed to the cercariae of *T. stagnicolae* for the first time and further suggest that repeated exposure is necessary to produce typical schistosome dermatitis.

9. *Notes on the Life Cycle of Schistocephalus sp., a Tapeworm from Gulls.* LYELL J. THOMAS, University of Illinois.

Shed segments of tapeworms were observed on the lake bed in about seven feet of water forming a ring around Pismier Island, Lake Michigan, June 7, 1947. Some were collected and proved to be those of a *Schistocephalus* sp. Three ring-bill gulls from this same island examined on July 13 had a total of five of these tapeworms. The worms were placed in a refrigerator for later experimental use. Ellipsoidal, operculate, dark amber colored eggs measuring $45\mu \times 87\mu$ were teased from the segments and placed in shallow water in Petri dishes to incubate at room temperature on July 2, 1947. Hatching proceeded with great speed when the fully embryonated eggs were exposed to strong lamp light. The coracidia measured $113\mu \times 147\mu$ and swam vigorously, without a rolling motion for four or five days before water taken in by the embryophore caused ciliary action to cease. Coracidia when exposed to both *Diaptomus* and *Cyclops* developed in *Cyclops leuckarti* and in no other species of *Cyclops*, in about fifteen days. Twelve guppies were placed with infected crustaceans on August 9 and three days later two of them died but were negative for tapeworms. Another large female guppy from this lot was examined September 29 but contained no tapeworms. The experiment is being continued.

10. *The Sedimentation Rate of Erythrocytes in Malaria Infections and Its Correlation with the Electrophoretic Mobility.* W. B. REDMOND AND W. S. POLLITZER, Emory University.

The sedimentation rate of red blood cells has been determined for normal pigeons, and for pigeons infected with *Plasmodium relictum*. During the period of increase in number of parasites the sedimentation rate changes little. Following the peak of parasitemia when the parasites are being destroyed, the sedimentation rate as well as the total amount of sedimentation increases 2 to 5 fold. When normal cells are placed in infected plasma there is no increase in the sedimentation rate, but when cells from an infected bird are suspended in normal plasma the rate is increased 6 to 8 fold. The total amount of sedimentation likewise increases to the same extent. One factor which appears to be of primary importance in determining the sedimentation is the volume per cent of packed cells. A decrease in volume of packed cells results in an increase in the sedimentation rate.

Although an increase in the serum globulins of the infected birds is found to be correlated with the increased sedimentation rate the plasma from an infected bird has no effect on the rate of normal cells.

A reduction in the electrophoretic mobility accompanies the increase in number of parasites. Tests have indicated that the reduction of the charge on the cells may be an important factor in the reduction of the sedimentation rate.

11. *Coccidia of the Mink.* NORMAN D. LEVINE, University of Illinois.

Four species of coccidia encountered in a dead mink (*Mustela vison*) from an Illinois fur farm are described: (1) *Eimeria vison* (oocysts ellipsoidal; mean oocyst dimensions 22.8 by 15.4 μ ; mean length-width index 1.50; no oocyst micropyle or residual body; large, coarsely granular sporocyst residual body); (2) *Eimeria mustelae* (oocysts subspherical; mean oocyst dimensions 16.1 by 14.4 μ ; mean length-width index 1.10; no oocyst micropyle or residual body; sporocyst residual body represented by a few scattered granules); (3) *Isospora bigemina* (oocyst wall very thin and stretched by sporocysts; mean oocyst dimensions 12.4 by 9.0 μ ; mean length-width index 1.4; no oocyst micropyle or residual body; large sporocyst residual body); and (4) *Isospora laidlawi* (oocysts ellipsoidal; mean oocyst dimensions 34.0 by 26.5 μ ; mean length-width index 1.26; no oocyst micropyle or residual body; small sporocyst residual body). *E. mustelae* and *I. laidlawi* are reported from the mink for the first time.

12. *The Effect of Sulfaquinoxaline on the Developmental Stages of Eimeria tenella.* ASHTON C. CUCKLER AND WALTHER H. OTT, Merck Institute for Therapeutic Research, Rahway, N. J.

Delaplane et al (1947, North Am. Vet., 28: 19-24) have found that sulfaquinoxaline (2-sulfanilamido quinoxaline) is highly effective for the control of cecal coccidiosis of the chicken. We are reporting studies on the mode of action of sulfaquinoxaline on the developmental stages of *Eimeria tenella*.

Equivalent groups of two-week-old chicks were placed on 0.1% drug diet one day before, concurrent with, or 1 to 7 days after, inoculation with 200,000 sporulated oocysts. At 24-hour

intervals after being placed on the drug diet, two chicks from the treated groups and a control were killed and the ceca were examined for progress of infection and therapeutic action. The ceca were preserved in Bouin's solution, sectioned and stained for study. On the tenth day the ceca of surviving chicks were homogenized and examined for oöcysts.

This study indicates that sulfaquinoxaline has a lethal effect on some of the sporozoites or first generation schizonts; however, the development of these stages is not completely inhibited as the large second generation schizonts produce merozoites and their liberation is accompanied by hemorrhage. Microscopically the second generation merozoites appear degenerated. Third generation schizonts, gametocytes, and oöcysts do not develop when chicks are placed on 0.1% sulfaquinoxaline drug diet before the fourth post-infection day.

13. *Observations on Leucocytozoon Infections in Birds Receiving Paludrine, Atebrin, and Sulphamerazine.* A. MURRAY FALLIS, Ontario Research Foundation, Toronto.

Paludrine, atebrin, and sulphamerazine were given to ducklings which were exposed to, or showing infection with *Leucocytozoon simondi*. Paludrine was given also to a crow showing a heavy infection of *L. sakharoffi*. The following dosages were used: One group of ducklings received 1 mg paludrine daily for 15 days following exposure; a second group received 1 mg paludrine t.i.d. for 4 days following appearance of infection in the blood; a third group received 2 mg atebrin daily for 15 days following exposure; a fourth group received 2 mg atebrin t.i.d. for 4 days following appearance of infection in the blood; a fifth group was given 50 mg sulphamerazine daily for 27 days following exposure. The drugs, in the dosages used, neither prevented nor cured infections with *L. simondi*, nor did they cause any obvious reduction in the number of parasites, and no morphological changes, which could be attributed to the drugs, were observed in the parasites. A crow which was heavily infected with *L. sakharoffi* was given 2 mg paludrine 4 times a day for 4 days but failed to recover. These results may be of interest in view of the action of these drugs on parasites of the related genus *Plasmodium*. The paludrine was kindly supplied by Imperial Chemical Industries.

14. *A New Ruling for the Ocular Disc of Special Usefulness in the Teaching of Medical Parasitology.* EDWARD P. OFFUTT, University of Rochester.

One of the greatest problems for the medical student in his consideration of parasitological specimens is to retain a concept of absolute size and of size relationships. To aid in this, and to facilitate the demonstration and study of specific points and areas, a new ruling for the ocular disc is suggested and a model demonstrated. Its features include: (1) a pointer; (2) an approximate ocular micrometer; (3) an indicator of red-blood-cell average size for the usual lenses of the student microscope; and, (4) a means for overlapping the field by a constant amount in a consistent manner, thereby permitting much easier systematic examination of entire preparations or specimens.

An opportunity will be given to express an interest in having this ruling made available on a commercial scale.

15. *The Metabolism of Mammalian Trypanosomes and Their Classification.* THEODOR VON BRAND AND ELEANOR JOHNSON TOBIE, National Institute of Health.

Hoare and Coutelen (1933) have divided the mammalian trypanosomes into Group A (*lewisii* group including *T. lewisii*, *T. theileri*, *T. melophagium*, and *T. cruzi*) and Group B with the subgroups *evansi* (*T. evansi*, *T. equinum*, and *T. equiperdum*), *vivax* (*T. vivax*, *T. caprae* and *T. uniforme*), *congolense* (*T. congolense* and *T. simiae*), and *brucei* (*T. brucei*, *T. rhodesiense*, and *T. gambiense*). This grouping which was based solely on morphological and developmental grounds, has apparently also a physiological basis. The members of Group A (*T. lewisii*, *T. conorhini*, and *T. cruzi*) have a cyanide-sensitive respiration and a relatively low rate of sugar consumption. All members of the *evansi* (*T. evansi*, *T. equinum*, and *T. equiperdum*), *congolense* (*T. congolense*), and *brucei* (*T. brucei*, *T. rhodesiense*, and *T. gambiense*) subgroups so far studied have a very high rate of sugar consumption. The respiration of the *evansi* and *brucei* subgroups is not inhibited by cyanide, that of the *congolense* subgroup is moderately cyanide-sensitive. No member of the *vivax* subgroup has so far been studied.

16. *Studies on Experimental Blackhead Infection in Turkeys.* FRANCIS MCKAY AND NEAL F. MOREHOUSE, Dr. Salisbury's Laboratories, Charles City, Iowa.

The use of embryonated ova of the cecal worm *Heterakis gallinae* for the transmission of the blackhead organism, *Histomonas meleagridis*, to turkeys, as reported by Graybill and Smith (1920), Tyzzer and Fabyan (1920) and Tyzzer (1934), has not been generally adopted by investigators working with this disease. In a series of 54 experiments involving 313 unmedicated turkey poults receiving embryonated *Heterakis gallinae* ova, 251 (80%) of them died of blackhead disease. A mortality of 100% was obtained in 24 of these experiments containing 113 poults, and at least 75% mortality was obtained in 43 of the 54 experiments. The *Heterakis*

gallinae ova used in these experiments were from cecal worms of chickens obtained from local dressing plants. Clinical symptoms of blackhead appeared as early as the 10th day after infection and as late as the 20th day after infection, most poult developing symptoms on the 12th to the 14th day. Mortality was greatest during the 15th to the 20th day after infection with deaths occurring as early as the 13th and as late as the 29th day.

17. *The Taxonomic Value of the Gnathosome in Certain Laelaptine Mites.* R. W. STRANDTMANN, University of Texas.

The difficulty frequently encountered by systematists in trying to place mites in their proper categories makes it imperative that all possible characters be examined for possible taxonomic values. The chelicerae, palpi, hypostome, epistome and also the tritosternum have long been recognized as of value in placing mites in the larger categories. In certain Laelaptine mites, however, these structures show variations which are of value on the genus and even species levels.

18. *The Resistance of Certain Adult Ducks to Infection by the Avian Malaria Parasite Plasmodium lophurae.* WILLIAM TRAGER, Rockefeller Institute for Medical Research.

White Pekin ducks 5 to 7 months old were inoculated intravenously with heparinized blood from young ducklings heavily infected with *P. lophurae*. The dosage was approximately one billion parasites per kilogram of body weight and the parasite count immediately after inoculation, was 50 to 200 parasites per 10,000 red blood cells. In most of the female ducks with active ovaries at the time of infection, as evidenced by actual egg production or by the finding of developing yolks at autopsy, the parasites underwent relatively little or no multiplication. All the females with inactive ovaries developed heavy infections, as did most of the males. The average free biotin and bound biotin-active lipoid material of the plasma before inoculation were highest in the egg-laying females, lowest in the males and intermediate in the females with inactive ovaries. In a few of the resistant individuals abnormal forms of the parasite appeared within a few days after inoculation and persisted for several weeks in small numbers.

19. *Immunity in Avian Malaria in Relation to Severity of Infection.* WENDELL GINGRICH, University of Texas.

In the course of testing drugs with *Plasmodium cathemerium* in canaries it has been observed frequently that infection is markedly suppressed but not cured, with no recrudescence or relapse following discontinuation of treatment. To test the immunity of animals with this type of infection 10 birds with very scant parasitemias (less than 1 to 18 parasites per 10,000 rbc) were superinfected with 10 million parasites after blood examination had become negative. Three showed no evidence of immunity, one doubtful, and 6 had about the same degree of immunity as birds which had recovered from typical acute infections.

20. *Progress Report on the Parasitic Stages of Chabertia ovina.* W. L. THRELKELD, Virginia Agricultural Experiment Station.

Parasite-free lambs, administered orally the infective larvae of *Chabertia ovina*, were necropsied on dates varying in length from the date of oral administration.

At 90 hours the third-stage larvae have undergone ecdysis and are either attached to the mucosa of the wall of the upper colon or have entered the wall. Petechial hemorrhages were profuse in the region and sections obtained therefrom revealed larvae attached to and embedded in the mucosa. The larvae measure 650 μ and the head region is characterized by lateral vacuolations. At 96 hours little change in larvae other than a slight thickening and a somewhat enlarged head region is discernible. At 6 days larvae average 1040 μ in length and a provisional buccal capsule is being formed. No sheaths could be determined. At 13 days the larvae measure 1300 μ in length the provisional buccal capsule is complete but no sheath could be determined and no sex differentiation is apparent. On the 18th day the male and female larvae are approximately the same length averaging slightly over 2 mm in length. Sex differentiation is evident and the permanent buccal capsule is being formed. The larvae are sheathed. This stage represents the later development of the 4th stage larvae as on the 25th day, ecdysis has taken place, the permanent buccal capsule is formed, but only one row of circum-oral teeth are present. The dorsal gutter has not yet split to form the sub-oral canal. On the 34th day the male and female are approximately the same length, averaging 7 mm, the sex organs being well developed. At 38 days copulation takes place and from 48 to 54 days adult worms are present and eggs appear in the feces.

21. *What Is a Virgula in Virgulate Cercariae?* FRANCIS J. KRUIDENIER, University of Michigan.

The virgula is a bilobed or paired organ with undetermined functions, in or near the oral sucker of certain species of xiphidiocercariae. In late larval and emerged cercariae, virgulae

are primarily paired structures, secondarily fused in some species, the reservoirs for secretions from paired unicellular glands arising antero-laterad to the acetabulum in close association with penetration glands. These paired pre-*virgula* glands empty through individual ducts into a forcibly enlarged weak oral sucker while still in the rediae. This secretion is readily hydrolyzed by inorganic acids or weak bases; a precipitate caused by acetic acid is hydrolyzable by inorganic acids. These reactions often indicate mucin compounds. Thionin, toluidin blue, and methylen blue demonstrated histochemically differential metachromatic mucin reactions of the *virgula* and glands as did neutral red, muchaematin, mucicarmine, and methyl green. Observations of emerged living and mounted specimens indicate that mucin discharged from the *virgula* supplements the weak oral sucker as a means of attachment. Published reports indicate further use during penetration into the host or during metacercarial development.

Paired glands parallel to those discussed above and others postero-lateral to the acetabulum discharge ventrally before the cercariae rupture the embryophore, enveloping them in a thin mucous sheath which is largely dissipated before cercariae emerge from snail hosts. Homologous glands and dispersed analogous cells are described in non-*virgulate* cercariae in abstract. . . . Apparently their mucous sheath aids and protects cercariae during their migrations through snail hosts.

22. *Quantitative Hookworm Diagnosis by Direct Smear.* PAUL C. BEAVER, Tulane University.

The direct fecal smear can be standardized by using a photoelectric type of light meter to measure its density (turbidity). Statistical analysis of multiple egg-count data indicates that eggs of the hookworm, *Necator americanus*, have random distribution in the fecal mass, and that on individual stools egg counts by the *standard direct smear (s.d.s.)* do not vary more than counts made by the Stoll dilution method. Close positive correlation exists between *s.d.s.* counts and dilution counts, the correlation being generally much closer when the latter are corrected to the formed stool basis. Day-to-day egg counts on the same individual are less variable by *s.d.s.* than by the dilution method. Counts by *s.d.s.* can be readily interpreted in terms of eggs per cc of formed stool or in terms of worm burden.

23. *A Method for Testing the Effect of Substances upon the Activity of the Larger Nematodes.* K. B. KERR AND J. W. CAVETT, Dr. Salsbury's Laboratories, Charles City, Iowa.

A procedure for the purpose of determining the effect of substances on the muscular activity of the larger nematodes is described. *Ascaridia galli*, has been used throughout this work. The worm is suspended in physiological saline at 40° C in a manner permitting a recording of its movements on a smoked drum. The saline is then removed, the test solution substituted, and its effect on the movements of the worm is recorded. The physical effect of changing solutions does not affect the worm, but care should be taken to have test solutions at approximately the same pH and temperature as the saline solution.

The effect of a number of known anthelmintics has been studied by this method and it has been found that these compounds cause a shortening of the nematode and a cessation of the normal irregular contractions demonstrated in the saline. Even after a considerable period of time there is seldom a recurrence of contractions, only a gradual relaxation of the worm. Upon removal from the apparatus the worms are usually still alive, for the release of tension permits them to curl and uncurl slowly. Substances which are not anthelmintics have no effect.

It is believed that this procedure provides a rapid and reasonably accurate method of determining whether a substance may have sufficient effect on the nematode to warrant an *in vivo* test for the anthelmintic effect of the substance.

24. *Immunity in Chickens to Ascaridia galli.* ELVIO H. SADUN, Johns Hopkins University.

In studies on resistance to *Ascaridia galli*, several groups of chickens have been experimentally infected with doses varying from 100 to 35,000 eggs. Out of 36 previously infected chickens, 30 eliminated all worms of the test dose while all controls, except one, harbored from 6 to 92 worms each. This acquired resistance persisted for over two months.

Sera from heavily infected chickens, tested *in vitro* with *Ascaridia* larvae, caused precipitates to be formed particularly around the mouth, thus indicating the presence of antibodies. No such precipitates were formed in normal sera. By serial dilutions it was possible to determine the titer of the antibodies responsible for the precipitates. Chickens which received repeated infections had a higher antibody titer than chickens with a single heavy infection. Older animals with an infection of 3,000 eggs, produced a higher antibody response than younger birds after receiving 12,000 eggs.

Intraperitoneal injection into 6-week-old chickens of serum from hyperimmune birds, elicited manifestations essentially the same as those seen in active immunity, showing that antibodies present in the serum are protective in nature and can be passively transferred. When very large

numbers of eggs were given to 3-day-old chickens, 100 per cent of those which received hyper-immune serum survived, while less than 50 per cent of those receiving normal serum or no serum at all, lived.

The various results indicate that natural age resistance and acquired immunity in chickens to *Ascaridia galli* are, partly at least, serological in nature.

25. *Factors in the Resistance of White Rats to Infection with the Cotton Rat Filarids.* J. ALLEN SCOTT, University of Texas.

By infecting white rats and cotton rats under parallel conditions certain factors explaining the relative insusceptibility of white rats have been identified. The number of worms recovered is less from white rats than from parallel cotton rats at early stages of development. The rate of growth is slower in white rats than in cotton rats. In the white rats more of the growing worms die before reaching maturity or are immobilized by a tissue reaction. One observation of apparent importance is that the tissue reaction not only involves dead worms, but it is often initiated by a covering of host cells surrounding portions of live worms which are active and unattached.

26. *Comparative Filaricidal Activity of Trivalent Arsenic and Antimony.* G. F. OTTO AND T. H. MAREN, Johns Hopkins University.

A number of amide substituted phenyl arsenoxides have been shown to have a marked lethal effect upon the adults of *Litomosoides carinii* *in vivo*; those which were further tested against the adults of *Dirofilaria immitis* were lethal to these organisms as well. The effects upon microfilaria are less clear. *In vitro* not only the amide substituted but other phenyl arsenoxides as well have an immediate lethal effect upon the *D. immitis* at much greater dilution than do any of the antimony compounds studied; however, *in vivo*, these have little or no demonstrable direct effect upon these microfilaria at several times doses required to kill the adult worms. On the other hand, while the *in vitro* activity of the several compounds against the microfilaria of *D. immitis* and *L. carinii* seems to be parallel, yet higher concentrations seemed to be needed to kill the latter. Conversely, however, the latter seem to respond more readily than the former *in vivo* while nevertheless being more resistant than the adults of either. It was not possible to study the *in vitro* action of these compounds upon the microfilaria of *Wuchereria bancrofti* but *in vivo* they are quickly killed by the same doses which have no demonstrable effect upon the microfilaria of *D. immitis* and *L. carinii*.

The trivalent antimonials, however, will kill microfilaria of all three species *in vitro* only at much greater concentrations than any of the trivalent arsenicals; furthermore, in our studies antimony killed these microfilaria much more slowly than did the arsenicals. Nevertheless, *in vivo* the trivalent antimony kills the microfilaria and injures the reproductive organs of the adult females without any appreciable number of the latter being killed; the injury to the reproductive tract is not immediately repaired if indeed it is not irreparable.

27. *Resumption of the Reproductive Phase of Trypanosoma lewisi in Recovered and Near Recovered Rats Following Both Reinoculation with the Parasite and Treatment with Sodium Salicylate.* ELERY R. BECKER AND MICHAEL G. LYSENKO, Iowa State College.

It has already been reported that the immunizing process which brings about the disappearance of the dividing forms of *Trypanosoma lewisi* from the blood is inhibited by sodium salicylate treatment. It did not succeed, however, to reinitiate the reproductive phase of the infection by such treatment of rats from adult trypanosomes surviving in the blood, or of recovered rats reinoculated with adult trypanosomes. In further experiments a certain number of such rats, i.e., either harboring adult trypanosomes or recovered, were subjected to salicylate treatment and reinoculated with 4-day trypanosomes, but infection did not follow. When, however, the donor rats as well as the acceptor rats were treated, infections characterized by the reproductive phase were re-established in a certain number of such rats. The reproductive phase persisted until drug-treatment was discontinued. In due time the flagellates transformed to the normal adult types which survived for a period, and finally disappeared from the blood.

28. *An Effective New Coccidiostatic.* PAUL D. HARWOOD, DOROTHY I. STUNZ, AND ROBERT WOLFGANG, Dr. Hess and Clark, Inc., Ashland, Ohio.

Furacin, which is manufactured by Eaton Laboratories, is chemically 5-nitro furfural semicarbazone. Since the manufacturers have demonstrated its bacteriostatic and bacteriocidal properties, it seemed advisable to test the compound against avian coccidiosis. 132 chicks, hatched July 6, 1947, were divided equally between 6 cages. One cage was kept as uninfected controls, but all the remaining chicks received by mouth 50,000 sporulated oöcysts of *Eimeria tenella* on July 24. Treatments were as follows: Cage 1, 0.011 per cent Furacin in the feed commencing July 22; cage 2, 0.011 per cent Furacin commencing July 26; cage 3, 0.017 per cent Furacin commencing July 26; cage 4, 0.017 per cent Furacin commencing July 27; cage 5, infected

controls; and cage 6, uninfected controls. The number of deaths occurring in each cage between July 24 and August 4 were as follows: Cage 1, 0; cage 2, 0; cage 3, 0; cage 4, 9; cage 5, 16; and cage 6, 0. When cages 4 and 5 are compared statistically, the value of chi-square is 4.78, which is well beyond the 5 per cent level. On August 9, all surviving chickens were given 200,000 sporulated oöcysts apiece. 5 birds from cage 6 died within the next two weeks. Throughout this experiment all birds which died were examined at necropsy, and all exhibited typical lesions of cecal coccidiosis. At these dosage levels, Furacin seems to be well tolerated, but at 0.1 per cent of the drug in the feed, chickens lose weight rapidly.

29. *A Method for Screening Anti-malarial Drugs in the Mosquito Host.* LEVON A. TERZIAN, Naval Medical Research Institute.

With a drug-diet method devised for the administration of various concentrations of anti-malarial drugs, it has been possible to drug mosquitoes infected with *Plasmodium gallinaceum* and to test the effects of these drugs on the sporogonous cycle of the parasite. It has been found that quinine, quinacrine and 7-chloro-4-(4-diethyl amino-1-methyl butyl amino)-3-methyl quinoline bisulfate (SN 6911) administered in maximum tolerated doses to infected mosquitoes has no effect on sporozoite production or sporozoite viability, and sporozoites from mosquitoes treated with these drugs, inoculated into normal chicks produce infections indistinguishable from infections produced by the inoculation of sporozoites from control mosquitoes maintained on sugar nutrient solutions alone. On the other hand, in infected mosquitoes treated with adequate drug concentrations of sodium sulfadiazine oöcysts fail to develop properly and sporozoites are produced only rarely, and those that are produced appear to be incapable of producing infection when inoculated into normal chicks. In each case inoculation of whole mosquitoes or several mosquito equivalents has failed to produce infection. Below a certain critical concentration of the drug, however, sporozoite production and viability are not affected and sporozoites from mosquitoes treated with lower drug concentrations produce characteristic infections. From the known drug effects of sodium sulfadiazine and the other drugs cited it would appear, therefore, that there is a definite relation between drug activity and effect in the sporozoite infected vertebrate host and drug activity in the infected invertebrate host. This method may offer further promise as another means for studies on parasite metabolism or for studies on the mechanisms of drug action.

30. *Observations on Oribatid Mites, Vectors of Moniezia expansa on Pastures, with a Report of Several New Vectors from the U. S.* K. C. KATES AND C. E. RUNKEL, U. S. Bureau of Animal Industry.

Collections of oribatid mites, concerned in transmission of *Moniezia expansa*, were made from (a) a large permanent sheep pasture at Beltsville, Md., (b) an irrigated pasture at Newell, South Dakota, and (c) a small pasture plot at Beltsville, Md., known to contain several species of oribatid mites, and "seeded" the previous year with *Moniezia* eggs in droppings. From these collections 6 species of oribatid mites, representing 5 genera and 4 families, contained cysticeroids of *M. expansa*. Only one of these species, *Galumna emarginatum*, is a known vector in the U. S. Another species, *Scheloribates laevigatus*, has been reported as a vector of moniezias and certain other anoplocephaline cestodes in Europe and Russia. Collections from pasture "(a)" revealed only one species, *Galumna virginiensis*, that contained cysticeroids; 34,224 mites examined, 3.9 per cent infected, 2,439 cysticeroids recovered. It is estimated that pasture "(a)" harbored in the top inch of turf a minimum of 6,000,000 *G. virginiensis*, containing more than 400,000 cysticeroids, per acre. *S. laevigatus* was the only infected mite from pasture "(b)"; 1,456 mites examined, 2.8 per cent infected, 64 cysticeroids recovered. Cysticeroids were found in 5 species of mites from "seeded" pasture plot "(c)"; *G. virginiensis*, 4,017 examined, 34 per cent infected, 3,873 cysticeroids recovered; *G. emarginatum*, 1,892 examined, 11 per cent infected, 378 cysticeroids recovered; *Oribatula minutus*, *Peloribates curtisii* and *Protoschelobates seggettii* (data combined), 866 examined, 6 per cent infected, 87 cysticeroids recovered. The largest number of cysticeroids obtained from one mite, a specimen of *G. virginiensis*, was 13.

31. *The Relationship between Trombiculid and Trombidiid Mites.* G. W. WHARTON, Duke University.

Until 1928 trombiculid mites were included in the family Trombidiidae. The trombiculids and trombidiids are similar in structure and habits. Both have larval stages that are parasitic and post-larval instars that are exclusively free-living. Trombiculids parasitize vertebrates; trombidiids parasitize invertebrates, usually insects. Collections of adults and the presumed larvae of a species of *Neotrombidium* in the Duke Forest has thrown some light on the relationships between the two families. The adults of *Neotrombidium* have recently been grouped with the trombiculids. The larvae of *Neotrombidium* have not been previously recognized. However they have been described under the generic name *Monunguis* and properly placed with the trombidiids

although they are morphologically similar to trombiculid larvae. It is possible therefore that the genus *Neotrombidium* represents a stage somewhat intermediate between the trombidid and trombiculid mites. *Neotrombidium* can best be grouped with the trombidids. The trombiculids most closely related to it are the members of the Leeuwenhoekinae. If the Leeuwenhoekinae represents the most primitive subfamily of the trombiculids, the trombiculinids and apoloninids must have arisen independently from them. The fourth subfamily, the Walchiinae, appears to have its origin from the Trombiculinae.

32. *Extended Persistence of Oöcysts of Plasmodium relictum in Culture.* GORDON H. BALL, University of California at Los Angeles.

Ball (1946, 1947) reported the persistence *in vitro* of oöcysts of *P. relictum* for six days on stomachs of *Culex tarsalis* cultured in hanging drops or in Carrel flasks. Failure of cultures to persist beyond this time was due in part to inability to maintain adequate nutritive and respiratory conditions with relatively small volumes of fluid. The successful culture of the erythrocyte stages of *P. knowlesi* by McKee, Ormsbee, Anfinsen, Geiman, and E. G. Ball (1946) with large volumes of fluid and with a continuous and controlled flow of gas offered a new method of attack. Infected mosquito stomachs were cultured on a cellophane membrane in a perfusion vessel, type 1. With this type of apparatus, apparently normal oöcysts of *P. relictum* could be demonstrated up to twenty-one days in culture. Despite better results in maintaining the oöcysts *in vitro*, there was little or no development of the oöcyst toward maturity.

33. *The Infectivity of Native Malarias in South Carolina to Anopheles quadrimaculatus.* MARTIN D. YOUNG, NEWTON F. HARDMAN, ROBERT W. BURGESS, WILLIAM C. FROHNE, AND CURTIS W. SABROSKY, U. S. Public Health Service.

In a field study in South Carolina, *Anopheles quadrimaculatus* mosquitoes were applied to native cases of malaria in Negroes to determine the infectivity of such malaria cases.

Of 142 feedings upon *Plasmodium falciparum* patients, 21 showed infections. Of 12,606 mosquitoes dissected, 2.5 per cent were infected. These infections developed normally in the mosquito, and in one out of two trials transmission was successful.

The majority of the mosquito infections occurred during the months of October through January. Mosquitoes kept at outside temperatures during October and November became infected.

One patient remained infective to mosquitoes over an eight-month period.

Of the 25 patients showing gametocytes, 60 per cent infected mosquitoes. The gametocyte densities resulting in infections were relatively low, none being over 90 per cmm. Thirteen of the infections resulted when the gametocytes were fewer than 10 per cmm.

Patients with asymptomatic parasitemias infected mosquitoes at about the same rate as those who were symptomatic. As the asymptomatic group is considerably larger than the other, it is concluded that this group is mainly responsible for the transmission of malaria.

There were 14 feedings upon *P. malariae*. Four cases showed gametocytes of a low density at feeding, and three lots of mosquitoes were infected. Of 586 mosquitoes dissected, 1.7 per cent were infected.

Of nine feedings upon *P. vivax*, none became infected, nor were gametocytes present.

It is concluded that the patient with an asymptomatic parasitemia, usually with a relatively low gametocyte density, is the important factor in the transmission and maintenance of *P. falciparum* and *P. malariae* malaria in the area studied.

34. *Expanding Horizons in the Recognition of a Phylum.* HARLEY J. VAN CLEAVE, University of Illinois.

Presidential Address.

35. *The Trematode Genus Tamerlania in Resident Birds of the United States.* ELON E. BYRD, University of Georgia.

During the past summer (1947) a large number of bird species from the regions about Highlands, North Carolina, and Mountain Lake, Virginia, were examined for internal parasites. Three of the species examined harbored flukes belonging to the genus *Tamerlania* Skrjabin, 1924, in the ureter. At Highlands, flukes of the genus were found in the Red-eyed Towhee, *Pipilo erythrophthalmus erythrophthalmus*, and the Parula Warbler, *Compsothlypis americana*; while the Red-eyed Towhee, *P. e. erythrophthalmus*, and the Oven-bird, *Sciurus aurocapillus*, from the region about Mountain Lake were parasitized by flukes of the genus. In so far as has been determined members of the genus *Tamerlania* have not been recorded from any of these three species of birds, nor has the genus been recorded previously from a species of bird known to be a permanent resident of the Continental United States. The presence of the genus in the Towhee constitutes such a new record. The only other record of the genus *Tamerlania* from a bird in the United States was reported by Penner (1939, J. Parasitol. 25: 421) from the migratory

Lincoln's Sparrow (Minnesota). Previously the genus has been recorded from various birds from Europe, Asia, Japan, Brazil, and Puerto Rico.

36. *Mucin in Developing Digenetic Trematodes*. FRANCIS J. KRUIDENIER, University of Michigan.

Anterior and posterior paired glands of virgulate cercariae (abstract 21) have homologues in widely different cercariae giving identical histochemical reactions. In some species structural adaptations conserve mucous secretions.

In developing microcercous cercariae of *Paragonimus kellicotti* (Troglorematidae) paired large anterior and posterior unicellular glands discharge mucus through individual ducts that empty respectively into an adherent antero-dorsal mass and a post-acetabular ventral groove. From the latter is discharged a mucus thread that anchors emerged cercariae to substrates. Analogous small cells provide emerging cercariae with mucinous envelopes. Cercariae of *Sellacotyle mustelae* (Troglorematidae) contain homologues.

A deep caudal pocket in cercariae of *Macroderoides typicus* (Macroderoididae) and paired, sub-terminal, bilateral pits in other cercariae of the Armatae and Ornatae groups open through channels dorsal to the tail base and act as reservoirs for paired posterior unicellular mucin glands that discharge before cercariae leave their rediae. Paired cells parallel penetration glands in position, development, and ducts and provide mucin for emerging cercariae.

Certain pleurophocercous cercariae (*Euryhalmis monorchis* and undetermined species) demonstrate paired anterior and posterior mucin glands during development but emerged cercariae retain no mucin reservoirs. Furocercous cercariae of *Clinostomum marginatum* have one pair of glands anterior to the eye spots with ducts paralleling those of penetration glands, and numerous mucin glands along the dorso-lateral surface. Certain glands discharge for use during emergence; five pairs are retained by emerged cercariae.

Wide occurrence in developing digenetic trematodes indicates that mucin must have considerable significance.

37. *Some Effects of Acanthocephala on the Large-Mouthed Black Bass*. CARL E. VENARD AND JOHN H. WARFEL, The Ohio State University and Western Reserve University.

In addition to other observations on the effects of *Leptorhynchoides thecatus* (Linton, 1891) and *Neoechinorhynchus cylindricus* (Van Cleave, 1913) on *Huro salmonides* (Lacépède), parasitized portions of alimentary canals were sectioned. Tissue damage due to the action of the spiny proboscis is evident in all of the material. There is mechanical destruction of much mucosa and submucosa. In one series of sections, where the hooks are deeply embedded in muscle tissue, the cells are cut as though by a sharp knife. The submucosa has many tunnels, often filled with cellular debris, and it is infiltrated with lymphocytes and many erythrocytes are present. Large masses of nuclei located around the parasites, or where the parasites had been, are evidence of the destruction of numerous cells. Aggregations of bacteria are present in some of the lesions.

38. *A Method for Relaxing and Fixing Large Cestodes*. ROBERT L. ROUDABUSH, Ward's Natural Science Establishment, Inc., Rochester, N. Y.

Tape worms, the size of *Taenia saginata*, are relaxed in water in an ice box over night. Specimens so relaxed are stretched to normal size by winding them on a beaker and fixed while still on the beaker thus avoiding contracted, distorted specimens. The formula for the fixing solution is 95% Ethyl Alcohol, 24 parts; Formalin, 15 parts; Glacial Acetic Acid, 5 parts; Glycerine, 10 parts; Tap Water, 46 parts.

39. *Mammalian Blood Flukes of Celebes*. ERNEST CARROLL FAUST, Tulane University, New Orleans, Louisiana, and C. BONNE, Universiteit van Indonesië, Batavia, Java.

Schistosoma japonicum was first found to be indigenous to the Lake Lindoë region of Celebes by Müller and Tesch (1937), who discovered human infection. The worm was later recovered from man, dog and deer from this area (Bonne et al, 1942). The molluscan host is thus far undiscovered. Fork-tailed cercariae, obtained from a species of *Lymnaea* in the vicinity of Lake Poso, were used experimentally in attempted infection of white rats and mice. On sacrifice the rats were negative but portal blood of the mice contained somewhat immature flukes. Study of the cercariae and adolescent worms indicates that they are not *S. japonicum* and possibly may belong to a different genus of Schistosomatidae.

40. *Germinal Masses in the Rediae of the Trematode Order Fasciolatoidea Seidat, 1936*. W. W. CORT, Johns Hopkins University, D. J. AMEEL, Kansas State College, and ANNE VAN DER WOUDE, University of Michigan.

The germinal material was studied in rediae of species of Paramphistomidae, Notocotylidae, Psilostomidae and Echinostomidae. In an amphistome, *Allassostoma parvum*, Stunkard, 1916,

and a notocotylid, *Quinqueserialis quinqueserialis* (Barker and Laughlin, 1911), very immature daughter rediae had at the posterior end of the body cavity simple morula-like germinal masses consisting of unicellular components. Evidence from the study of older rediae indicated that these simple germinal masses were gradually used up by the formation of embryos from their components.

Studies were also made of the germinal material in the rediae of a psilostome, *Psilostomum ondatrae* Price, 1931, and of certain echinostome species, *Echinostoma revolutum* (Froelich, 1802), *Petasiger nitidus* Linton, 1928 and at least two that were unidentified. In the very youngest daughter rediae the germinal masses also had only unicellular components. But in older rediae, both mother and daughter, larger germinal masses were present with both unicellular and multicellular components. These were frequently attached at one side of the wall of the posterior end of the body cavity, and were still present in the very oldest rediae examined. They resemble in structure the floating germinal masses of the daughter sporocysts of the plagiurchiids.

Perhaps the simpler germinal masses in the amphistome and notocotylid rediae represent a more primitive condition associated with a rather limited production of individuals, while the large persistent germinal masses of the psilostome and the echinostome rediae are an adaptation for greater embryo production over a longer period of time.

41. *Germinal Material in Mother Sporocysts and Rediae of Halipegus eccentricus* Thomas, 1939. D. J. AMEEL, Kansas State College, W. W. CORT, Johns Hopkins University, and ANNE VAN DER WOUDE, University of Michigan.

Halipegus eccentricus, a hemiurid trematode from the eustachian tubes of the frog, has its mother sporocyst and one generation of rediae in species of *Physa* in the Douglas Lake region, Michigan. A very unusual development of germinal masses was observed in a large series of immature stages of mother sporocysts and rediae from both experimental and natural infections. A morula-like germinal mass consisting only of unicellular components is present in the miracidium and youngest mother sporocyst. In a stage 0.1 mm in length, the body cavity is almost completely filled with germinal masses composed of both unicellular and multicellular components. In later stages these produce large numbers of embryos, and in almost mature mother sporocysts all the germinal masses have broken down into embryos except one attached near the posterior end. This was still present in the oldest mother sporocyst seen. It is estimated that by this germinal mechanism in the mother sporocyst 200 or more redial embryos can be produced. In very immature redial embryos a morula-like germinal mass of unicellular components is present near the posterior end. In older embryos germinal masses are found all along the body cavity. In later stages these break up into cercarial embryos, leaving finally a single large germinal mass in the posterior end which is still present and giving off embryos in mature rediae. This mechanism provides for the production of the large numbers of cercarial embryos that are present in the large mature rediae of this species.

42. *Porocephalus crotali* Humbolt (*Pentastomida*) in Oklahoma. J. TEAGUE SELF, University of Oklahoma.

Rattlesnakes examined from Oklahoma show an over-all eight per cent infection of the pentastomid *Porocephalus crotali* Humbolt. Heaviest infections occur in specimens isolated in the areas of hibernation caves in the Wichita Mountains Wildlife Refuge. As many as 158 specimens have been collected from the lung of one individual.

Specimens show size relationship with the three species of the genus, *P. crotali*, *P. clavatus* and *P. stilesi*, and differ from previous generic designations in the absence of auxiliary hooks and the sameness of lateral and median hooks. Further study may require a revision of the generic description or the establishment of a new genus.

The deer mouse, *Peromyscus leucopus aridulus* Osgood, serves as the principal secondary host, particularly around hibernation caves.

43. *Additional Infection Experiments with the Hookworm, Bunostomum phlebotomum, in the Calf.* ROY L. MAYHEW, Louisiana State University.

The following experiments are a continuation of those reported by the writer in the Journal of Parasitology 2: 14 Supplement. Five additional calves have been inoculated by placing infective hookworm larvae on the skin. Eggs were recovered from the manure of four of the animals between the 52nd and the 62nd day after inoculation. No eggs were recovered from the fifth even though he showed the most severe symptoms. Three animals had periods of diarrhea on the following days after inoculation—from the 16th to 26th, 28th to 62nd, and from the 37th to the 53rd. Weights were taken over a period of 125 days following inoculation. An examination of the weights showed that four animals had more or less irregularity in their gains during the period between 29 and 60 days following inoculation. During the two months following the end of the pre-patent period all animals made regular gains in weight. One un-

inoculated animal kept under the same conditions of management made regular gains during the entire experiment. One calf died on the 53rd day after a period of diarrhea lasting 17 days and a loss of 58 pounds.

These data suggest the following conclusions: (1) That the hookworm of cattle is the cause of severe symptoms and considerable economic loss to the cattlemen, (2) that infection takes place through the skin thus making barn and shade sanitation a necessary part of the management program, (3) that the symptoms of infection are developed during the pre-patent period, and (4) that a general improvement in condition follows during the adult life of the parasite.

These observations strongly suggest that treatment is of little value during the period of acute symptoms but should, however, be utilized in the program of control to eliminate the egg-producing adults and thus reduce the number of larvae about the premises which may serve as a cause of further infections.

44. *On Certain Pseudophyllidean Cestodes from Alaskan Pinnipeds.* HORACE W. STUNKARD, New York University.

Tapeworms from the bearded seal, *Phoca barbata*, the fur seal, *Callorhinus ursinus*, and the sea lion, *Eumetopias jubata*, have been studied. There are 3 and perhaps 4 species represented. The worms from *P. barbata* are monogonadic; those from *C. ursinus* represent 2 species, one monogonadic, the other diplogonadic; while those from *E. jubata* are similar to and possibly identical with the diplogonadic form from *C. ursinus*. The observed differences may be the result of development in different hosts. Similar and probably in part identical tapeworms from the bearded and fur seals were reported by Wardle, McLeod, and Stewart (1947, *J. Parasitol.* 33: 319-330). The worms were regarded as congeneric and a new genus, *Cordicephalus*, with *Taenia phocarum* Fabricius, 1780 as genotype, was erected to contain them. All diphyllbothrid cestodes from members of Phocidae in North Atlantic waters were assigned to a single species, *C. phocarus* (Fabricius, 1780) and those from members of the Otariidae to *C. arctocephalinus* (Johnston, 1937). It should be noted that *T. phocarum* Fabricius, 1780 was designated as type of a new genus, *Pyramicocephalus* by Monticelli, 1890; consequently *Cordicephalus* is a synonym of *Pyramicocephalus*. Although the allocation of species by Wardle et al is not acceptable, specific determination of the specimens at hand must be tentative until the morphology and taxonomic status of previously described species are clarified.

45. *Nature of Duodenal Nematode Growth-Inhibiting Factor.* LYMAN P. FRICK AND J. E. ACKERT, Kansas State College.

In studies interrupted by World War II, it was demonstrated that duodenal mucus of growing chickens contains a nematode-growth-inhibiting factor. *Ascaridia galli* removed from fowls and placed in a nutrient solution grow in length. When duodenal mucus from resistant chickens is placed in the nutrient worm culture the worms' growth is retarded, sometimes to the extent of losing length. It was to determine if such effects on the worms are permanent that the studies here reported were made.

Three groups of *A. galli* subjected to the action of the mucus for a week, were remeasured and transferred to the regular nutrient solution. The worms in two of the three groups, after losing length in the mucus cultures, had the ability to recover and resume growth when they were transferred to the nutrient solution. In the next test with three groups of *A. galli*, all of the worms lost length in the mucus culture and resumed growth when they were returned to the nutrient solution.

The results of these tests indicate that the inhibitory nematode growth factor is nutritional in character, and that its effects are temporary.

46. *Further Studies on Hydrogen-ion Concentration as a Factor in Age Resistance to the Fowl Ascarid.* B. B. RIEDEL AND J. E. ACKERT, Kansas State College.

Hydrogen-ion concentration of the habitat of the fowl nematode *Ascaridia galli* as a possible factor in age resistance was mentioned in this body in 1945 when comparisons were made between 4-week and 13-week fowls. The studies, which have been continued, cover various regions of the alimentary tract of chickens up to 35 weeks of age.

Normal White Leghorn chickens reach the maximum resistance to the *Ascaridia* when they are about 17 weeks old. Had the study ceased with chickens 11 weeks old, it would have appeared that hydrogen-ion concentration might be a factor in reducing the growth rate of the *Ascaridia*, as the average pH reading at that age of chicken (6.09) was markedly lower than that at the 4-week age (6.44). As the older fowls were tested, however, the pH readings veered toward those of the chickens of the susceptible ages. For example, resistant chickens 17 weeks of age averaged 6.67 as compared with 6.51 for susceptible fowls 8 weeks old. The fact that the worm habitat in chickens 17 to 35 weeks of age had about the same hydrogen-ion concentration as did that of the young birds 1 to 8 weeks of age tends to remove hydrogen-ion concentration as a factor in age resistance of chickens to the *Ascaridia*.

No constant trend in hydrogen-ion concentration with age of fowls was observed for the following organs: proventriculus, gizzard, anterior duodenum, posterior duodenum (worm habitat), posterior intestine (at yolk sac diverticulum), and caecum (11 and 15 weeks).

47. *Age of Ascaridia Egg Culture a Factor in Worm Growth.* J. E. ACKERT, RHODA M. COOPER, AND L. W. DEWHIRST, Kansas State College.

Variability in results in experimental parasitology is one of the problems of the investigator. A number of possible factors in variability of the number and size of *Ascaridia galli* in chickens have been studied; but apparently not age of egg culture.

To test age of egg culture as a possible factor in variability, 1000 embryonated eggs of *Ascaridia galli* were taken at random from an old egg culture (120 days) and given in 100-egg lots to 10 young chickens. From a young egg culture (36 days), 1000 eggs were similarly taken and fed to 10 other young chickens of the same age.

Three weeks later, the test was terminated and the worms from each chicken collected, counted and measured. The results showed that the worms from the chickens fed from the old egg culture averaged 3.4 worms per fowl as compared with 10.2 worms from the young egg culture. As to growth, the worms from the old egg culture averaged 15.5 mm in length as compared with an average worm length of 23.6 mm from the young egg culture, a significant difference.

These results indicate that to avoid needless variability in size of worms, embryonated eggs for comparative infection experiments should be taken from egg cultures of approximately the same age.

48. *The Status of the Sodium Fluoride Treatment for the Removal of Large Roundworms from Swine.* A. O. FOSTER, F. D. ENZIE, R. T. HABERMANN, AND R. W. ALLEN, U. S. Bureau of Animal Industry.

In consequence of a search for an improved treatment for the removal of large roundworms from swine, sodium fluoride, data on which were first published in July, 1945, has been found to be highly efficient in removing both ascarids and stomach worms, well tolerated when given in therapeutic dosages in dry, ground feed, and easy to administer. The treatment has been extensively investigated in the United States and Australia, and has received wide field use. It is replacing to an increasing extent the commonly employed swine ascaricides of recent date, namely, oil of chenopodium and phenothiazine, over each of which the sodium fluoride treatment possesses measurable net superiority when judged by the obvious criteria of safety, efficacy, ease of administration, and cost. The method of use that appears most satisfactory consists in the administration of the chemical in dry, ground feed, at a concentration of 1 per cent for a period of 1 day. While the effective dose rate is in the vicinity of 0.1 gram per pound of live weight, both critical investigation and clinical field experience have indicated that there is risk of intoxication if the chemical is administered in capsules or drenches, or in garbage, slops, or milk. The treatment is contraindicated in swine exhibiting symptoms of gastroenteritis. Rarely does the treatment give rise to symptoms other than occasional vomiting and softening of the feces.

49. *Screening Large Numbers of New Chemical Compounds for Anthelmintic Activity Using Infections of Nippostrongylus muris in mice.* STERLING BRACKETT AND ALEXANDER BLIZNICK, American Cyanamid Company, Stamford, Connecticut.

Factors of importance in selecting a parasite-host combination for use in large-scale screening of new compounds for leads to anthelmintic activity are: (1) ease of producing standardized experimental infections in large numbers of test animals, (2) smallness of host in order to conserve drug which is usually available in quantities of only a few grams in the case of new compounds, (3) availability of a uniform strain of host in large numbers at low cost, (4) ease of caring for and dosing host, (5) an easily read endpoint, and (6) reproducibility of results. Experimentally induced infections with *Nippostrongylus muris* in young mice (10-14 g) using worm counts at autopsy as the endpoint, have been found to meet to a satisfactory extent all of the above requirements, and permits a team of two persons to screen-test from 40 to 80 new compounds a week using as little as 0.3 g of each compound. Greatest emphasis has been on drug-diet method of drug administration. Standard anthelmintics such as hexylresorcinol, phenothiazine, β -naphthol, santonin, gentian violet and arecoline, which are not too volatile to be tested in this way, are ineffective. The only compounds found to have unquestionable activity thus far, namely trichloroacetamide and related compounds, only affect the larval stages migrating in the tissues. Whether these or other compounds which may be found active against *N. muris* in the future will have practical application can be determined only by trials against parasites of economic importance and in the natural host.

50. *Further Studies on the Maintenance of Cultures of Endamoeba histolytica without Viable Bacteria.* LEON JACOBS, National Institute of Health.

In an earlier paper (Am. J. Hyg., Sept. 1947) use of penicillin for elimination of viable bacteria from cultures of *Endamoeba histolytica* was described. Maintenance for over 3 months of one series of amoeba cultures after elimination of viable bacteria, and establishment of another series were reported. Since this report the second series was maintained for 6 months and two other series have been established.

In establishing cultures the medium used is Locke's-egg conditioned by growth of *Escherichia coli* for 24 hours after which the bacteria are heat-killed. This medium is inoculated from cultures of amoebae with *Clostridium perfringens*, and penicillin added to eliminate the viable bacteria introduced. Usually multiplication of amoebae is obtained through 2 or 3 transplants at 48 hours in such tubes. In later transplants the amoebae usually become scarce. Survival rarely follows but the series described have been derived from the few tubes in which it has occurred. After elimination of *C. perfringens*, penicillin is not needed. Cultures without viable bacteria show few amoebae, but multiplication does occur since a large series of cultures has been produced from a few tubes.

The rarity of survivors through penicillin treatment suggests that the successes obtained may have been due to acclimatization of amoebae to the insufficient medium. The penicillin technique has been exploited in hope of discovering accessory materials required by the amoebae. Dead yeast, yeast juices, ether-killed bacteria, and plasmotyzates of bacteria have been tried as supplements without success.

51. *Studies on the Growth Requirements of Metacystic Endamoeba histolytica.* CHARLES W. REES and LUCY V. REARDON, National Institute of Health.

Metacystic amoebae produced in microcultures from cysts of *Endamoeba histolytica* freed from bacteria by microisolation were studied in various kinds of egg media enriched with B vitamins and cholesterol, and in various wholly liquid media, including trypsin digests of egg, combinations of amino acids, extracts of liver and beef, filtrates of bacterial cultures, and suspensions of heat-killed bacteria. The metacystic amoebae failed to divide although they lived in some cases up to 96 hours. Inoculation of the microcultures with organism *t* resulted in division and limited growth of the amoebae. Transfer, by means of a microsyringe, of the bacteria-free metacystic amoebae to stock tubes of whole egg medium inoculated with organism *t* has regularly produced cultures of amoebae. Improvements in the technique of microisolation have led to the practice of isolating 100 cysts per microculture with the production of up to 40 bacteria-free metacystic amoebae per culture. Experimental work on growth requirements of the amoebae has thus been facilitated since in previous work not more than 25 cysts per culture were isolated.

52. *Culture Experiments on Intestinal Flagellates. IV. Selected Longevity Records to September 15, 1947.* D. H. WENRICH, University of Pennsylvania.

Employing the methods previously described, the life of flagellates in single culture tubes was prolonged by adding food (usually powdered gastric mucin or Loeffler's dried blood serum) as needed, and distilled water to maintain fluid level. For any culture reported below which was referred to in a table of a previous paper, the designation used there is indicated in parentheses. Longevity figures are in days.

(1) *Trichomonas hominis* from man (6A), 495; (2) *T. wenyoni* from white rat (Rat 1A, 1C), 965; (3) *T. wenyoni* from hamster (Ham. AR/2b), 1245; (4) same from a man who swallowed a culture (Ha/Ho 1B), 1185; (5) *T. batrachorum* from frog (*Rana pipiens*) (5C/1a), 1750 (nearly 5 years); (6) *T. prowazeki* from *Amphiuma means*, 546; (7) *Monocercomonoides* sp. from guinea-pig (G.-p. 2A), 683; (8) *M. sp.* from Japanese beetle larva (Jap. 3C), 882; (9) *M. sp.* from tipulid larvae (Tip. 3A, 3B), 872; (10) *M. sp.* from lizard (*Sceloporus* sp.), 331; (11) *Monocercomonas (Eutrichomastix)* sp. from same lizard, 331; (12) *Trimitus parva* from frog (*Rana pipiens*) (11A), 937; (13) *Hexamitus* sp. from bullfrog (*Rana catesbeiana*), 520. A new strain of *Trichomonas angusta* from *Rana pipiens* is being maintained in one-half strength Ringer's with Loeffler's dried blood serum and rice starch; longest survival to date in any one tube, 130 days.

No. 5 was started in water; all others in modified or in diluted Ringer's. No. 1 was maintained at 32–33° C; all others at room temperatures. All but nos. 5 and 7 are still positive.

53. *The Phase Microscope in the Examination of Endamoeba.* ERNEST HARTMAN and GEORGE J. SCHEFF, The Chicago Medical School. (Also by demonstration.)

The use of the phase microscope permits the direct observation of internal structure of living *Endamoeba* free from the distortion of fixed material. The nuclei are prominent and show denser bodies somewhat different from the appearance of iron hematoxylin prepared material.

54. *The Fate of Various Species of Trypanosomes in Triatoma.* A. PACKCHANIAN, The University of Texas.

Laboratory-reared, normal *Triatoma gerstaeckeri* were allowed to feed on guinea pigs infected with *Trypanosoma gambiense* and *Trypanosoma rhodesiense*. Likewise, normal *Triatoma* were permitted to feed on rats experimentally infected with *T. lewisi*. Other batches of *Triatoma* were fed on mice (*Mus musculus*) infected with *T. duttoni*.

None of the *T. gambiense*, *T. rhodesiense*, *T. lewisi*, or *T. duttoni* multiplied in *Triatoma*. The trypanosomes survived a few days in the insect guts, their numbers gradually diminishing; at the end of ten days no viable flagellates were noted, and the susceptible animal inoculation tests with crushed *Triatoma* were all negative.

T. gambiense, *T. rhodesiense*, and *T. lewisi* were passed also into soft ticks (*Ornithodoros turicata*). None of these micro-organisms multiplied or survived in the ticks and at the end of about ten days susceptible animal inoculation tests were likewise negative for trypanosomes.

* 55. *Studies of Sheep Parasites. VIII. Overwintering of Nematode Larvae.* PHILIP A. HAWKINS AND MOACYR GOMES DE FREITAS, Michigan State College.

During a five-year period it was noted that there was a decided decrease in nematode egg counts, and an increase in concentration of hemoglobin during the late winter months in ewes and yearling lambs. In late February or March there were increased egg counts and a lowering of hemoglobin. It was assumed that this was produced by the development of infective larvae in the warmer environment of the barn where the sheep were kept. During the winter of 1946-47 examinations were made of the straw bedding in the sheep barn at regular intervals by a modified Baermann technique. No infective larvae were recovered between December 16, 1946 and April 7, 1947. Small numbers of infective larvae of *Nematodirus* sp., *Haemonchus contortus*, and other short tail larvae probably *Trichostrongylus* sp. were recovered on April 14, 1946. In one yearling ewe that died February 31, 1947, a few immature specimens of *H. contortus* were recovered. No immature nematodes were recovered from two yearlings dying February 21 and February 24, respectively. One lamb born March 7 died April 10 of pneumonia and at autopsy revealed 10 *Nematodirus* sp. and 1 *Trichostrongylus* sp.

Between early December and February 10, the mean temperatures in the straw bedding were usually below freezing. The week of February 10-17, the mean temperature was 38° F, February 17-24, it was 35° F and during March it was usually not below 40° F. Therefore under natural conditions of infection a mean temperature of approximately 40° F is required for the development of infective larvae. The increase in egg counts in late April is due to the ingestion of larvae during that month, but increases during the winter months are not correlated with the presence of infective larvae. These increases are referable to variations in the rate of egg production of the female worms which pass over the winter in the animal. The causes of this variation are unknown.

56. *The Effect of Aralen on Giardia lamblia Infections in Children.* J. C. SWARTZWELDER AND T. C. PAPERMASTER, Louisiana State University.

In view of reported cures of giardiasis with Aralen diphosphate, the following observations are recorded. Five children infected with *Giardia lamblia* were treated with Chloroquine diphosphate (Aralen, SN7618) to determine whether or not this new antimalarial compound, like atabrine, was also effective against that intestinal flagellate. Prior to treatment, stool examinations in these cases showed large numbers of *G. lamblia* cysts in direct fecal smears. Both direct smear and zinc sulphate centrifugal flotation technics were employed during and following therapy to determine the presence or absence of the parasites in the stools. The dosages of Aralen employed and the approximate weight of the individuals treated were as follows: Cases 1, 2 and 5—35 to 40 lbs.—dosage 125 mg ($\frac{1}{2}$ tablet) of Aralen diphosphate twice daily for three days; case 3—26 lbs.—62 mg ($\frac{1}{4}$ tablet) daily for 4 days; and case 4—30 lbs.—62 mg of drug daily for 3 days. No significant untoward reactions to the drug were observed clinically in the treated children. With the above dosage, there was a temporary disappearance or a reduction in number of *Giardia* cysts in the stool in four of the five cases following treatment. However, the parasites reappeared or persisted in the stool following a brief interval of apparent absence or depression. No reduction was noted in one case. These results do not preclude the possibility that higher dosages of Aralen, as used for adults, may be more effective against *G. lamblia* infections. Repeated stool examinations over an extended period after treatment are necessary to evaluate the permanent effect of the drug on *Giardia* infections.

57. *Bite of Amblyomma americanum Associated with Possible Tick Paralysis.* J. C. SWARTZWELDER AND J. H. SEABURY, Louisiana State University.

Two partially fed nymphs of *Amblyomma americanum*, the lone star tick, were found attached to the skin in the mid-thoracic region over the spine in a white, male child, 12 years of age. The boy was admitted to Charity Hospital of Louisiana at New Orleans, with a provisional diagnosis of poliomyelitis in September, 1946, between two and three days after the sudden onset

of his illness. Clinical manifestations and physical findings included a staggering shuffling gait, weakness of the lower extremities, fever (103.6° F), pharyngitis, headache, vomiting, nuchal stiffness, diminished patellar reflexes, diminished left abdominal reflex, slight tenderness of thigh and calf muscles and of the abdomen, and moderate inguinal and cervical lymphadenopathy. Spinal fluid examination revealed no increase in cell count. The patient improved rapidly subsequent to removal of the ticks. Examination of the ticks revealed that they were members of the genus *Amblyomma*, therefore they were submitted to a specialist for further study. G. M. Kohls of the Rocky Mountain Laboratory identified the specimens as partially fed nymphs of *Amblyomma americanum*. In view of the fact that this species has not previously been associated with tick paralysis, the case is reported. Although the possibility of the diagnosis of abortive poliomyelitis can not absolutely be excluded in this case, the occurrence of two partially fed ticks attached to the skin over the spine and the fact that rapid and spontaneous recovery occurred subsequent to removal of the ticks, appears to be more than coincidental.

58. *Report of Ten Cases of Balantidium coli Infection.* J. C. SWARTZWELDER, Louisiana State University.

In 1910, a fatal case of balantidiasis was reported from Charity Hospital at New Orleans. Since then at least 10 additional infections with *Balantidium coli* have been recorded at that institution. Nine of the ten previously unreported cases have been observed since 1936 and three were diagnosed in 1947. The patients ranged in age from 2 to 65 years. Five were fifteen years or older. Eight of the individuals were white and 9 were males. Five of the adults were farmers. All of the patients, except possibly one, resided in rural areas. Families of two patients, who were questioned on the subject, raised hogs. Eight of the patients complained of diarrhea, the most common symptom. Five individuals noted blood in their stools. Several individuals complained of weight loss. Diagnosis was made by stool examination, proctoscopy, and in one case the ciliate was detected in a section of appendix. Proctoscopy was done by attending physicians in at least six cases as a diagnostic measure, to study the lesions, or to observe the effect of therapy. The latter technic was a valuable supplement to stool examination. In only one case did the writer observe cysts of *Balantidium coli*, and these were accompanied by trophozoites, the usual diagnostic stage. Seven of the cases also had concomitant infections with two or more helminths, namely *Ascaris lumbricoides*, *Trichuris trichiura*, *Necator americanus*, and *Enterobius vermicularis*. Spontaneous temporary disappearance of the ciliate necessitates multiple stool examination for diagnosis and study over an extended period for evaluation of therapy.

59. *The Life History and Bionomics of Heterakis bonasae a Caecal Nematode of Bobwhite Quail, Colinus virginianus and C. virginianus texanus.* J. W. WARD, University of Mississippi.

The life history of *Heterakis bonasae* has been demonstrated to be direct. Day-old white leghorn chicks were utilized as the definitive hosts. Sixteen days is the average time for the undifferentiated eggs to develop into the infective larvae. Forty-seven days is the average time for development from infective larvae to mature worms. The eggs show a resistance to prolonged drought, moisture and low temperature. Undifferentiated eggs were found to develop normally after standing in ice for thirty days. Repeated wetting and drying had no effect upon the viability of undifferentiated eggs. Eggs developed to normal infective larvae after drying for thirteen months.

Young worms were found to burrow into the mucosa at the junction of the small intestine and the caecae.

60. *Further Taxonomic Studies on Internal Parasites of Horses and Mules.* J. W. WARD, University of Mississippi.

Results of this study are based upon the findings at autopsy of twenty-four horses and mules. The age of the animals varied from fifteen days to twenty years or more. Age of animals and parasite load could not be correlated. Animals three weeks of age were found to be heavily parasitized with three or more nematodes. Fourteen genera and nineteen species of parasites were recorded, as follows: *Gastrophilus nasalis*; *G. intestinalis*; *Parascaris equorum*; *Strongylus equinus*; *S. vulgaris*; *S. edentatus*; *Triodontophorus minor*; *T. serratus*; *Cylicocercus goldi*; *Cylicocycylus elongatus*; *C. nassatus*; *Oxyurus equi*; *Dictyocaulus arnfieldi*; *Setaria equina*; *Strongyloides* sp.; *Anoplocephala* sp.; *Trichonema* sp.; *Habronema* sp.; *Gongylonema* sp.

61. *Studies on Parasites and Food Habits of Foxes.* J. W. WARD, University of Mississippi.

The results of this study are based upon the findings at autopsy of seventeen wild gray foxes collected by trapping from the vicinity of Pontotoc and Springville, Mississippi. Three animals were examined for external parasites. Fifteen of the seventeen animals, or 88%, were found to be infected with the cestode *Taenia pisiformis*. The number of individual scoleces per

host varied from one to fifty-four. The average number being thirteen. Two foxes, or 12%, were infected with *Toxacara canis*. One animal was infected with *Uncinaria stenocephala*. Two external parasites were recovered from one animal, namely, *Cediopsyllus simplex* and *Dermacentor variabilis*. Stomach contents showed the common food to be field rats.

62. *Longevity and Rates of Infectivity of the Free-Swimming Miracidia of Schistosoma mansoni*. JOSÉ F. MALDONADO AND J. ACOSTA-MATIENZO, School of Tropical Medicine, San Juan, Puerto Rico.

During the study of the larval cycle of *S. mansoni* in *Australorbis glabratus*, several problems of epidemiologic importance arose. The first which has occupied our minds is the question of how long do the free-swimming miracidia of this parasite live and to what extent are they capable of penetrating into the snail with increasing age. Normal, recently passed eggs, obtained from the same patient throughout the experiment, were isolated singly in rain-water in watch glasses and continuously observed under the dissecting microscope, to determine their exact time of hatching. From the moment of hatching and thereafter at exact hourly intervals, a small *A. glabratus* was exposed to each miracidium and whether penetration occurred was determined. Slightly over 100 miracidia were used for each exposure interval.

The data show that the longevity of the free-swimming miracidium of *S. mansoni* is eight to nine hours. Seventy per cent of the miracidia were able to penetrate into the snail host immediately after hatching. The percentage of survival and successful penetration were as follows: one hour, 65.4; 2 hours, 65.0; 3 hours, 52.0; 4 hours, 35.9; 5 hours, 36.9; 6 hours, 11.5; 7 hours, 10.8; 8 hours, 3.9. At nine hours none of the miracidia were alive. Hatchability, mortality and infectivity varied markedly from day to day.

A negligible number of eggs was able to hatch beyond the 48th hour after the stools were put in contact with water.

63. *An Investigation of Some Possible Sources of Trichina Infection in a Central Illinois Community*. WAYNE W. WANTLAND AND PAUL MARTIN, Illinois Wesleyan University.

A study was made of thirty-seven hogs, thirty-two rats, and ten samples of pork products. A small packing plant, two farms where garbage is fed to hogs, three city dumps and local stores were the sources of material. Three methods of examination were used: (1) direct microscopic study consisting of clamping muscle tissue between two glass slides and examining with compound microscope, (2) a modified Baermann digestion method and microscopic examination of the residue following centrifugation, and (3) microscopic examination of intestinal contents of rats. No adults or larvae of *Trichinella spiralis* were found.

64. *The Localization of Adult Trichinella spiralis in the Intestinal Tract of Young and Old Mice*. JOHN E. LARSH, JR., AND JAMES R. HENDRICKS, University of North Carolina.

Using old rats for class study it has been noted that the majority of *T. spiralis* adults are found in the first half of the small intestine. When the reverse was found to be the case in young rats, it suggested an interesting relationship worthy of further study.

White mice were used in four similar experiments completed thus far. A total of 27 young mice (4 to 6 weeks) and 26 old mice (12 to 16 weeks) were used. Each animal was infected with 300 *T. spiralis* larvae and counts were made of adults found five days later. The results of all four experiments were similar. The young mice averaged 20% development of adults and of this number (60) 36% were found in the first half of the small intestine and 64% in the second half. The old mice averaged 30% development of adult worms and of the total (91) 84.7% were found in the first half of the small intestine and 15.3% in the second half. This showed a striking difference, therefore, in the location of adult *T. spiralis* in the intestinal tract of young and old mice. The intestinal emptying time of young mice is much greater than that of old mice, so that it may be a factor involved in this phenomenon. Studies are in progress using drugs, at present morphine sulphate, to test this possibility by artificially slowing the intestinal emptying time of young mice after infection.

65. *The Effect of Thyroid and Thiouracil on the Natural Resistance of Mice to Infection with Hymenolepis*. JOHN E. LARSH, JR., University of North Carolina.

The mice used were divided into five groups. Those of group A were five months old and were given each day by mouth 2 mg of thyroid extract. Group B mice of the same age served as untreated controls for group A. The mice of group C were one month old and each was given 2 mg of thiouracil (American Cyanamid Company). Those of group D were the same age but untreated. The last group, 2½ months old, included as controls on age resistance, was injected with normal saline. After one month of treatment the animals were infected. Accounting for the age of the animals of the various groups, there was little difference noted in the percentage development of *Hymenolepis*. The food intake likewise was similar for all groups.

In a repeat of this experiment, grouping the mice as above and using 3 mg of thyroid and 2.2 mg of thiouracil, a great difference was noted. The thyroid animals showed a much higher percentage development than their controls of the same age, whereas all of the thiouracil animals showed about the same rate of development. The food intake of the thyroid mice was higher than for their controls, whereas there was little difference noted in the thiouracil groups. There is indication, therefore, that thyroid given to old mice has an adverse effect upon normal age resistance. It is well known that hyperthyroidism produces a drain on the vitamin stores of the body. Therefore, these animals may have exhibited decreased age resistance to *Hymenolepis* as a result of vitamin deficiencies, which have already been shown to be involved in alcoholic debilitation of mice infected with this parasite.

66. *The White Rat and Guinea Pig as Hosts for the Larvae of the Brown Dog Tick*, *Rhipicephalus sanguineus*. GEORGE W. LUTTERMOSER, University of Pennsylvania.

An attempt was made to rear the different stages of the life cycle of the brown dog tick, *Rhipicephalus sanguineus*, with white mice, white rats and guinea pigs as hosts. Eggs deposited by female ticks which had previously engorged on dogs, were placed in Mason pint jars with tops of fine copper screen (475 mesh per square inch) held in place by standard jar rings. The jars were stored in a saturated atmosphere of a small humidity chamber. At room temperature the eggs hatched in about six weeks. At least 100 larval ticks were then transferred to an animal which had been previously isolated in a battery jar partially filled with shavings and surrounded by oil. Larvae were collected from the sides of the jar, from the shavings or from each animal during a period of a week and were returned to the Mason jars and humidity chamber for observation of further development. The nymphs were transferred to the animals maintained under the same conditions as described above. Unengorged nymphs and adult *R. sanguineus* were collected from dogs and were handled in the same fashion as the larvae.

About 5% of the larvae of *R. sanguineus* fed on guinea pigs in three tests, and they engorged on shaven white rats in each of the two tests made. The engorged larvae recovered from both the rats and guinea pigs molted and were again placed on these animals, but they failed to engorge. The nymphal and adult ticks collected from dogs likewise failed to engorge on the rodents. The larvae did not engorge on four unshaven white rats or on a white mouse. These data would seem to indicate that *R. sanguineus* can adapt itself to a life cycle with more than one host.

67. *Susceptibility and Immunity of Marine Fishes to Benedenia (= Epibdella) melleni (MacCallum), a Monogenetic Trematode. III. Natural Hosts in the West Indies*. ROSS F. NIGRELLI, New York Aquarium, New York Zoological Society.

Benedenia melleni was originally described in 1927 from captive marine fishes in the New York Aquarium where a large variety of spiny-rayed species was found susceptible. Some can develop a partial or total immunity; others are always immune. Infection spreads rapidly to all susceptible fishes kept in a closed circulation, constantly recirculating the same water. Death from heavy infestation often occurs. A survey of *Benedenia* infestations among fishes of Bimini, B.W.I., showed the following to be naturally infected, but never with more than three of the parasites: *Epinephelus striatus*, *Lutjanus apodus*, *Chaetodon ocellatus*, *C. striatus*, *C. capistratus*, *Pomacanthus arcuatus*, *P. paru*, *Holacanthus tricolor* and *H. ciliaris*. These are species that become most heavily parasitized in captivity. The following fishes are known to become infected in captivity, but were found free of parasites at Bimini: *Holocentrus ascensionis*, *Caranx* spp., *Haemulon* spp., *Acanthurus caeruleus*, *A. hepatus*, *Balistes vetula*, *Lactophrys bicaudatus* and *Diodon hystrix*. The following species, kept in a large, open-work, stockade-like enclosure and a concrete tank with open "windows," off the dock at Bimini, were heavily parasitized: *trachinotus goodiei*, *C. ocellatus*, *S. striatus*, *P. arcuatus*, *P. paru*, *H. ciliaris*, *Melichthys piceus* and *Alutera scripta*. An unlimited supply of fresh sea-water does not prevent heavy infestations. Proximity to infected fishes or to possible resting places for developing eggs of the parasite are apparently the critical factors.

68. *Parasites of the Ranidae (Amphibia)*. I. A. C. WALTON, Knox College.

The Ranidae are primarily found in the Eastern Hemisphere, particularly in Africa and the tropical and subtropical Orient. One genus (*Rana*) is cosmopolitan in distribution. Among the *Arthroplectinae* (Africa) the following hosts and parasites have been recorded: 1. *Arthroplectis minutus*—*Endotrombicula penetrans* (Acarina); and 2. *A. ogoensis*—*Cylindrotaenia americana* (Cestoda). 1. *Phrynobatrachus graueri*—*Aplectana congolense* (Nematoda), and a larval mite of Loveridge, 1929 (Acarina); and 2. *P. natalensis*—*Balantidium entozoon* and *Opalina natalensis* (Protozoa), and *Schöngastia pillarsi* (Acarina). Among the *Astylosterninae* (Africa) the records are: *Astylosternum robustum*—*Africanus astylosterni*, *Amplicaeum novempapillatum*, and *Oxysomatium chameleonis* (Nematoda). The *Petropedetinae* (Africa) record

is: *Arthroleptides dutoiti*—*Aplectana* sp.? of Loveridge, 1926 (Nematoda). Records for members of the *Raninae* (Cosmopolitan) are as follows: *Ooiedozyga lima* (China)—*Diplodiscus japonicus*, larval *Encyclometra asymmetrica*, and larval *Pharyngostomum cordatum* (Trematoda). 1. *Rana acquipicalata* (Mozambique)—*Cylindrotaenia ?americana* (Cestoda). 2. *R. adspersa* (Africa)—*Agamascaris africana* (larval Nematoda); *Ophiotaenia schultzei* (Cestoda); and *Dactylosoma* sp.? of Wenyon, 1931, *Hexamita ranae*, and *Protoopalina mossambicensis* (Protozoa). 3. *R. aesopus* (U.S.A.)—*Cosmocercoides dukae*, *Foleyella* sp.? of Causey, 1938, and larval *Ophidascaris labiatopapillosa* (Nematoda); and *Brachycoelium salamandrae* and *Megalodiscus temperatus* (Trematoda). 4. *R. agilis* (Europe)—*Nematotaenia dispar* (Cestoda); and *Acanthocephalus ranae* (Acanthocephala). 5. *R. albilabris* (Africa)—larval *Filaria* sp.? of Schwetz, 1930 (Nematoda); and *Dactylosoma ranarum* and *Trypanosoma rotatorium* (Protozoa). 6. *R. amurensis* (Siberia)—*Haematoloechus schultzei* and *H. sibericus* (Trematoda). 7. *R. angolensis* (Africa)—*Haemogregarina theileri*, *Trypanosoma nelspruitense*, *T. rotatorium* and *T. spp.*? of Laveran, 1904, and Laveran & Mesnil, 1912 (Protozoa). 8. *R. angulosa* (China)—*Glythelminis staffordi* (Trematoda). 9. *R. areolata* (U.S.A.)—*Cosmocercoides dukae* (Nematoda); *Haplometrana utahensis* and *Megalodiscus temperatus* (Trematoda); and *Balanitidium* sp.? of Metcalf, 1923, and *Opalina kennicotti* (Protozoa).

69. *Parasites of the Ranidae (Amphibia). II.* A. C. WALTON, Knox College.

10. *Rana arvalis* (Europe)—*Aplectana acuminata*, *Cosmocerca commutata*, *Oswaldocruzia filiformis*, *Oxysomatium brevicaudatum* and *Rhabdias bufonis* (Nematoda); *Brandesia turgida*, larval *Distoma* sp.? of Tiniofeev, 1900, *Gorgoderina cygnoidea*, *Gorgoderina vitelliloba*, *Haplometra cylindracea*, *Monostoma ellipticum*, *Opisthioglyphe endoloba*, *Opisthodiscus subclavatus*, *Pleurogenes claviger*, *P. medians*, *Polystoma integerrimum* and *Prostocus confusus* (Trematoda); and *Acanthocephalus ranae* (Acanthocephala). 11. *R. arv. altaica* (Siberia)—*Haematoloechus sibericus* and *Pleurogenes intermedius* (Trematoda). 12. *R. arv. isaitschikovi* (Europe)—*Aplectana acuminata*, *Cosmocerca ornata*, *Oswaldocruzia filiformis*, *Oxysomatium brevicaudatum*, *O. longispiculum* and *Rhabdias bufonis* (Nematoda); and *Gorgoderina vitelliloba*, *Haplometra cylindracea* and *Opisthioglyphe histrix* (Trematoda). 13. *R. aurora* (U.S.A.)—*Cosmocercoides dukae*, *Oswaldocruzia waltoni* and *Rhabdias joaquinensis* (Nematoda); *Glythelminis californiensis*, *Gorgoderina aurora*, *G. multilobata*, *Lecithodendrium lynchi* and (in tadpoles) larval *Paralechthorichis syntomentera* (Trematoda); *Crepidobothrium olor* (Cestoda); and *Opalina obtrigonoidea lata* (Protozoa). 14. *R. aur. draytoni* (U.S.A.)—*Spirooura pretiosa* (Nematoda); *Cephalogonimus brevicirrus*, larval *Clinostomum* sp.? of Ingles, 1936, *Glythelminis californiensis*, *Haematoloechus kernensis*, *H. oxyorchis* (exper.) and *H. humidus* (Trematoda); and *Hexamita batrachorum*, *H. ovata*, *Opalina draytonii*, *Trichomonas augusta*, *Trypanosoma* sp.? of Wenrich, 1935, and *Zelleriella ranaxena* (Protozoa). 15. *R. boylii* (U.S.A.)—larval *Clinostomum* sp.? of Ingles, 1936, *Glythelminis californiensis*, *Gorgoderina multilobata*, *Haematoloechus butensis*, *H. oxyorchis* (exper.) and *Haliplus aspinus* (Trematoda); Paruterine larvae of Ingles, 1936 (Cestoda); and *Hexamita batrachorum*, *H. ovata*, *Opalina virguloidea* and *Trichomonas augusta* (Protozoa). 16. *R. boylii sierrae* (U.S.A.)—*Spirooura pretiosa* and *S. ranae* (Nematoda). 17. *R. capita* (?=*R. aesopus*) (U.S.A.)—*Opalina obtrigonoidea lata* (Protozoa). 18. *R. cantabrigensis* (U.S.A.)—*Megalodiscus temperatus* (Trematoda). 19. *R. cancrivora* (Asia)—*Kiricephalus pattoni* (Porocephalida).

70. *Parasites of the Ranidae (Amphibia). III.* A. C. WALTON, Knox College.

20. *Rana catesbiana* (N. Amer.)—larval *Abbreviata ranae*, larval *Agamascaris odontocephala*, *Cosmocercoides dukae*, larval *Dujardinia* sp.? of Brandt, 1936, larval *Eustrongylides wenrichi*, *Foleyella ranae*, larval *Icosiella quadrutuberculata*, *Multicaecum tenuicollis*, *Ophidascaris labiatopapillosa*, *Oswaldocruzia collaris*, *O. leidy*, *Oxysomatium americana*, *Rhabdias ranae*, *Spinitectus* sp.? of Trowbridge & Hefley, 1934, *Spirooura catesbiana*, "*Spiroptera*" *mugientis* and *Strongylurus ranae* (Nematoda); larval *Alaria mustelae*, *Allasostoma parva*, larval Amphistome of Trowbridge & Hefley, 1934, larval *Aurodistomum chelydrae* (exper. in tadpoles), *Cephalogonimus americanus* (?=*C. retusus*), *Cercaria vesiculosa*, larval *Clinostomum complanatum*, larval *Dasyatra conferta* (in tadpoles), larval *D. villicaeca* (in tadpoles), *Glythelminis queta*, *G. subtropica*, *Gorgoderina amplicava*, *G. cygnoidea*, *G. minima*, *Gorgoderina attenuata*, *G. simplex*, *G. vitelliloba*, *Gyrodactylus* sp.? of Stunkard & Dunihue, 1933 (on tadpoles), *Haematoloechus brevilexus*, *H. complexus*, *H. floedae*, *H. longiplexus*, *H. parviplexus*, *H. variegatus*, *H. varioplexus*, *Haliplus amherstensis*, *H. eccentricus*, *H. occiduus*, *Loxogenes arcanum*, *Megalodiscus intermedius*, *M. temperatus*, larval *Neoreneifer aniarum* (in tadpoles) and *Opisthodiscus ?subclavatus* (Trematoda); *Cylindrotaenia americana*, *Ophiotaenia magna*, *O. saphena* and *Proteocephalid* cysts of Brandt, 1936 (Cestoda); larval *Centrorhynchus* sp.? of Brandt, 1936 (Acanthocephala); *Amphileptus branchiarum*, *Cytamoeba bacterifera*, *Discophrya* sp.? of Metcalf, 1923, *Entamoeba ranarum*, *Epistylis* sp.? of Wenrich, 1935 (on tadpoles),

Haemogregarina catesbiana, *Hexamita intestinalis*, *Ichthyobodo necatrix* (in tadpoles), *Karyolysus* sp.? of Brandt, 1936, *Lankesterella canadensis* L. spp.? of Brandt, 1936, and Wenyon, 1933, *Leptotheca ohlmacheri*, *Mastigamoeba hylae*, *Nyctotherus fecundiformis*, *Opalina larvarum* (in tadpoles), *Opercularis* sp.? of Wenrich, 1935 (on tadpoles), *Plasmodium catesbiana*, *Retortamonas dobelli*, *Rhabdostyla* sp.? of Wenrich, 1935 (on tadpoles), *Spirochaeta manitoui*, *Trichomonas augusta*, *T. batrachorum*, *T.* sp.? of Thompson, 1934, *Trichodina* spp.? of Diller, 1928, and Wenrich, (on tadpoles), *Trypanosoma inopinatum*, *T. rotatorium*, *T.* sp.? of Hegner, 1920, and *Vorticella* sp.? of Wenrich, 1935 (on tadpoles) (Protozoa); *Macrobella dictetra* (Annelida); *Hannemania penetrans* (Acarina); and *Culex territans* (Diptera).

Symposium on Exoerythrocytic Forms of Malarial Parasites

Program

71. Introduction to Symposium. CLAY G. HUFF, Naval Medical Research Institute.

Some of the historical background for the following five papers on exoerythrocytic forms of malarial parasites is given. In addition, the relationship between these stages and other malaria-like organisms and between the avian and hypothetical human forms is discussed. An attempt is made to orient the subjects presented in the five papers and to indicate some of the conclusions that can be drawn as well as the limitations imposed by our lack of knowledge.

72. A Search for the Pre-erythrocytic Stages of *Plasmodium vivax* and of *P. cynomolgi*. CLAY G. HUFF, Naval Medical Research Institute, AND FREDERICK COULSTON, E. I. du Pont de Nemours and Company.

Using methods which were successful for finding pre-erythrocytic stages of four species of avian malaria an intensive search was made for the corresponding stages of *P. vivax* in human volunteers. Salivary glands from infected *Anopheles quadrimaculatus* were inoculated in minimum amounts of fluid into the skin, lymph nodes, muscle and isolated veins of volunteers in the Manteno State Hospital and the Stateville Penitentiary (Illinois). The inoculated area was then biopsied, fixed, sectioned, and stained by methods described in our earlier papers. Although remnants of the inoculated material were usually located in the sections no pre-erythrocytic stages were discovered. Material from 24 biopsied areas of skin, 7 biopsied lymph nodes, 4 isolated veins, and 1 muscle biopsy were examined. The intervals at which the biopsies were taken extended from 2 to 150 hours but the majority were made at 24, 48, and 72 hours. In attempts to demonstrate the presence of viable parasites in the inoculated areas by subinoculation these areas were biopsied and subinoculated or grafted into other volunteers. Infections developed in all of the original recipients of the sporozoites but none developed in volunteers receiving the biopsied areas of skin. Since pre-erythrocytic stages of some avian malariae will develop in hosts which do not develop parasitemia, attempts were made to find similar stages in the sites of inoculation of sporozoites of *P. vivax* in liver, spleen, bone marrow, and skin of five species of monkeys (*Macaca mulata*, *Cercopithecus aethiops sabaeus*, *Cercopithecus a. pygerythrus*, *Cercocebus fuliginosus*, and *Papio papio*). The findings were negative. Similar experiments with *P. cynomolgi* and *Macaca mulata* at approximately 24, 48, and 72 hour intervals in liver, spleen, bone marrow, and skin also yielded negative findings.

73. The Chemotherapy of Malaria in Relation to Our Knowledge of Exoerythrocytic Forms. G. ROBERT COATNEY AND W. CLARK COOPER, National Institute of Health.

In this paper, the authors attempt to bring together all information relating to the action of drugs upon the exoerythrocytic forms in malaria. Emphasis is given to the direct proof that the sulfonamides and certain other compounds destroy these forms in *P. gallinaceum* malaria. The indirect evidence obtained from chemotherapeutic studies relative to the occurrence and nature of exoerythrocytic forms in the higher vertebrates is critically discussed.

74. The Chemotherapy and Immunology of Pre-erythrocytic Stages in Avian Malaria. FREDERICK COULSTON, E. I. du Pont de Nemours and Company, AND CLAY G. HUFF, Naval Medical Research Institute.

With the development of a technique whereby the tissue stages of avian malarial parasites could be studied, it was possible to consider immunological and chemotherapeutic aspects of the pre-erythrocytic stages. In general, the visible effects on the parasite and the host seem to be similar, whether chemotherapy is employed or the natural and acquired immunity of the host is allowed to follow the normal course of events. Elsewhere, the authors have considered such important factors in the immunology of avian malaria as the normal and abnormal host and the tissue-blood barrier. This work will be reviewed and integrated.

Chemotherapeutic agents of a prophylactic or suppressive type were given to chickens in-

fected with sporozoites of *P. gallinaceum*. The cryptozoites and metacryptozoites of these birds were studied by histological methods. Blood levels of the drugs were obtained whenever possible.

Taken as a whole, no drug tested was capable of preventing the development of the cryptozoites. However, certain drugs, such as sulfadiazine, effectively prevented the growth of the succeeding generations of metacryptozoites, whereas quinine and quinidine did not prevent their development. An important consideration is the fact that all good antimalarial drugs tested exhibited some effect on the metacryptozoites.

75. RICHARD JANVIER PORTER, University of Michigan.
(Invited Paper; No Abstract Received.)

76. *Response of Exo-erythrocytic Forms to Alterations in the Life-Cycle of Plasmodium gallinaceum.* VICTOR H. HAAS, AIMEE WILCOX, R. L. LAIRD, FRANCES MOORE EWING, AND NELL COLEMAN, National Institute of Health and University of Tennessee.

It is known that in *Plasmodium gallinaceum* passed alternately through mosquitoes and chicks, exo-erythrocytic forms attain their greatest density near the end of the prepatent period, while in passage by blood inoculation alone, these forms become prevalent after parasitemia has subsided.

This report describes another type of host-parasite relationship, in which profuse growth of exo-erythrocytic forms kills the host before erythrocytic parasites become numerous. This phenomenon occurs: (a) In chick embryos infected by mosquitoes. (b) In chicks and chick embryos inoculated with brain emulsions containing exo-erythrocytic forms. In embryos, the accompanying few erythrocytic forms produce no pigment. These infections maintain their characteristics in both chicks and chick embryos through serial brain emulsion inoculations, but are convertible by 3 or 4 blood passages to the classical blood inoculation type. In such converted strains, gametocytes are extremely rare, but low-grade mosquito infections are obtainable. (c) In chicks inoculated with sporozoites from mosquitoes previously infected on chicks of the blood-inoculated series.

This type of host-parasite relationship, accompanying alterations in the life cycle of *P. gallinaceum*, represents a response on the part of the exo-erythrocytic forms which could not generally occur under natural conditions, since death of the host before erythrocytic forms become numerous would prevent cyclical development of the parasite.

AMERICAN SOCIETY OF PARASITOLOGISTS
CONSTITUTION

(Including revisions of December 30, 1931, December 28, 1932, December 29, 1933, December 31, 1940, December 30, 1941, and December 27, 1946.)

NAME AND OBJECT

The name of the society is the American Society of Parasitologists.

The object of the society is the association of workers in the field of Parasitology for the presentation and discussion of new or important facts and problems in that science and for the adoption of such measures as will tend to the advancement of parasitological teaching and investigation in this country.

MEMBERSHIP

The members of the society shall be of two classes, active and foreign honorary.

Any person with suitable educational qualification interested in parasitology may be a candidate for active membership.

Any foreign scientist who has made eminent contributions to Parasitology may be eligible for honorary membership.

Candidates for membership shall be elected by the Council.

OFFICERS

The officers of the society shall be a President and a Vice-President, who shall be elected for one year; a Secretary and a Treasurer, who shall be elected for two years; and members at large of the Council.

The Council shall consist of the President, the Vice-President, the Secretary, the Treasurer, the Chairman of the Editorial Committee, and eight members elected by ballot from the society at large, two for four years, two for three years, two for two years and two for one year. After the first year two members at large of the Council shall be elected each year to serve four years.

If any vacancy occurs among the officers, the Council is authorized to appoint a member to fill out the unexpired portion of the current year.

The routine business of the society shall be administered by the Council.

Five shall constitute a quorum of the Council.

DUES

The dues, to include subscription to the JOURNAL OF PARASITOLOGY, shall be four dollars the year for active members, unless changed by vote of the society.

MEETINGS

There shall be an annual meeting and such other scientific or business meetings as the Council shall determine.

During the annual meeting a business meeting will be held for the election of officers for the ensuing year and for the transaction of other business.

JOURNAL OF PARASITOLOGY

This Journal, the property of the society, is its official organ. Responsibility for its conduct shall rest with the Council which shall select the editorial staff and set the price of subscription.

ENDOWMENT FUND

Provision is made for the establishment of a permanent endowment fund, the principal of which may be expended only by a three-fourths vote of all members of the Council and approval by a three-fourths vote of the members of the society present at a regular meeting. The Council shall be entrusted with the maintenance of the fund, and the use of the income therefrom.

AMENDMENT

On recommendation of a two-thirds vote of the Council, the constitution may be amended by a two-thirds vote of the members present at any regular business meeting of the society, provided that at least 30 days' notice has been given to the membership of the proposed amendment.

BY-LAWS

MEMBERSHIP

1. *Election of Members.* An affirmative vote of all Council members present at a meeting shall be necessary for the election of candidates for membership. If the vote is taken by mail ballot, an affirmative vote of all members of the Council replying within thirty days shall be required.

2. *Members in Good Standing.* Members in good standing are those whose current dues are paid.

3. *Delinquent Members.* The JOURNAL OF PARASITOLOGY shall not be sent to members in arrears, and members in arrears for the current year and two previous years shall be dropped from the roll of the society at the end of the current year.

4. *Reinstatement of Delinquent Members.* Members dropped for non-payment of dues, or who have resigned, may be reinstated by the payment of all dues in arrears. Otherwise, the applicant must apply for election as a new member.

5. *Non-subscribing Members.* Where a second membership in the society is taken in the same immediate family, the second member may join upon the payment of annual dues of one dollar, but without receiving the JOURNAL OF PARASITOLOGY.

6. *Honorary Life Members.* Upon unanimous vote, the Council may recommend that the society confer honorary life membership upon distinguished American parasitologists over sixty years of age. Honorary Life Members shall enjoy full membership privileges and shall be exempted from the payment of dues. The number of Honorary Life Members at any one time shall be limited to five.

7. *Foreign Honorary Members.* Upon unanimous vote, the Council at its annual meeting may elect Foreign Honorary Members. No two men from the same country shall be elected in the same year. The number of Foreign Honorary Members at any one time shall be limited to twelve. Foreign Honorary Members may receive the JOURNAL OF PARASITOLOGY upon payment of membership dues.

PRESENTATION OF PAPERS AT ANNUAL MEETINGS

1. Except for invited papers, and papers coming to the American Society of Parasitologists program through joint sessions with another society or section, all persons presenting papers must be members of the society in good standing, or must be introduced by a member in good standing.

2. Each member of the society in good standing may be allotted not more than fifteen minutes of program time, to be used in person or by a non-member introduced by him. If the program is crowded, the maximal allotment of time may be reduced to ten minutes.

3. Papers offered for presentation too late to be included on the printed program may be presented at the conclusion of any of the scientific sessions provided that the presiding officer obtains the consent of the members present.

OFFICERS

1. The Council shall act as a nominating committee and at the annual meeting shall submit to the society at least one nominee for each office to be filled. The Secretary shall invite the members of the society to submit nominations for consideration by the Council.

2. The Treasurer of the society shall act also as Treasurer for the JOURNAL OF PARASITOLOGY and shall be bonded for the sum of two thousand dollars.

3. A sum of fifty dollars shall be allotted annually to the Secretary, to the Treasurer, and to the Chairman of the Editorial Committee for attendance at the annual meeting.

MANAGEMENT OF THE JOURNAL OF PARASITOLOGY

1. The JOURNAL OF PARASITOLOGY, as the official organ of the society, shall be managed by an Editorial Committee appointed by the Council for a five-year period, and shall consist of one Protozoologist, one Helminthologist, and one Entomologist, one of whom shall be appointed by the Council to act as Chairman.

2. The Editorial Committee shall be assisted by an Editorial Board consisting of twelve members appointed by the Council for a four-year period in such a way that three members will retire and three new members shall be elected each year. These shall be elected on the basis of attainment, interest in the Society, geographical location, and representation of the various fields of the science.

3. The price of the JOURNAL OF PARASITOLOGY shall be five dollars per volume, except to members of the American Society of Parasitologists who shall receive it as a membership privilege included in the annual dues of four dollars.

ENDOWMENT FUND

1. Council shall select a Custodian of the Endowment Fund and two associates to whom it may delegate responsibility for management of the fund. The Custodian shall make an annual accounting to Council and such other reports as Council may request. The approval of two of the three custodians shall be necessary for the purchase, sale or exchange of securities. One of the three custodians shall be the Treasurer of the society and his signature shall be required on all vouchers of expenditure from the fund.

ADDITIONS AND AMENDMENTS

1. Additional by-laws may be created by a two-thirds vote of Council members present at a meeting, or by an affirmative vote of nine Council members in a ballot conducted by mail. By the same procedure existing by-laws may be repealed, amended or suspended.

AMERICAN SOCIETY OF PARASITOLOGISTS

Officers for 1947

HARLEY J. VAN CLEAVE, University of Illinois	President
CLAY G. HUFF, Naval Medical Research Institute	Vice-President
JAMES T. CULBERTSON, University of Arkansas	Secretary
ROBERT M. STABLER, Colorado College	Treasurer

Council Member Ex Officio¹

HORACE W. STUNKARD, New York University	Chairman, Editorial Committee
---	-------------------------------

*Council Members at Large**(with date of expiration of term)*

1950	EMMETT W. PRICE, U. S. Bureau of Animal Industry
1950	MARTIN D. YOUNG, U. S. Public Health Service
1949	WILLARD H. WRIGHT, U. S. Public Health Service
1949	THOMAS W. M. CAMERON, McGill University
1948	GILBERT F. OTTO, Johns Hopkins University
1948	G. ROBERT COATNEY, National Institute of Health
1947	HAROLD W. BROWN, Columbia University
1947	CORNELIUS B. PHILIP, U. S. Public Health Service

*Representatives of the Society on the Council of the American
Association for the Advancement of Science
(2-year terms expire 1947)*

G. ROBERT COATNEY	WILLARD H. WRIGHT
-------------------	-------------------

*Representatives of the Society on the Council of the
Union of American Biological Societies
(3-year terms expire 1948)*

DONALD L. AUGUSTINE	ROBERT M. STABLER
---------------------	-------------------

Editorial Committee of the Journal of Parasitology

HORACE W. STUNKARD, Chairman	to serve until 1948
WILLIAM A. RILEY	to serve until 1948
DAVID H. WENRICH	to serve until 1948

Editorial Board of the Journal of Parasitology

1950	RICHARD J. PORTER, University of Michigan
1950	WILLIAM TRAGER, Rockefeller Institute for Medical Research
1950	ARTHUR C. WALTON, Knox College
1949	LLOYD E. ROZEBOOM, Johns Hopkins University
1949	WILLIAM W. CORT, Johns Hopkins University
1949	RAYMOND M. CABLE, Purdue University
1948	LLOYD A. SPINDLER, U. S. Bureau of Animal Industry
1948	CHARLES W. REES, National Institute of Health
1948	WILLIAM L. JELLISON, U. S. Public Health Service
1947	LOWELL T. COGGESHALL, University of Michigan
1947	JOHN T. LUCKER, U. S. Bureau of Animal Industry
1947	NORMAN R. STOLL, Rockefeller Institute for Medical Research

List of Former Officers

<i>President</i>	<i>Vice-President</i>
1925 HENRY B. WARD*	SAMUEL T. DARLING*
1926 CHARLES W. STILES*	CHARLES A. KOFOID*
1927 RICHARD P. STRONG	EDWIN LINTON*
1928 CHARLES A. KOFOID*	ROBERT HEGNER*
1929 NATHAN A. COBB*	GEORGE R. LARUE
1930 WILLIAM W. CORT	ERNEST CARROLL FAUST

¹ Beginning in 1942, the Chairman, Editorial Committee, became ex officio member of Council.

* Deceased.

1931	WILLIAM A. RILEY	ASA C. CHANDLER
1932	MAURICE C. HALL*	WILLIAM H. TALIAFERRO
1933	WILLIAM H. TALIAFERRO	FRED C. BISHOPP
1934	ERNEST E. TYZZER	JAMES C. ACKERT
1935	CHARLES F. CRAIG	HARLEY J. VAN CLEAVE
1936	ROBERT HEGNER*	WILLIAM B. HERMS
1937	GEORGE R. LARUE	DAVID H. WENRICH
1938	FRED C. BISHOPP	ELERY R. BECKER
1939	HORACE W. STUNKARD	HENRY E. MELENEY
1940	DAVID H. WENRICH	GOTTHOLD STEINER
1941	JAMES E. ACKERT	JUSTIN ANDREWS
1942	HENRY E. MELENEY	RUDOLF W. GLASER*
1943	HENRY E. MELENEY	RUDOLF W. GLASER*
1944	HENRY E. EWING	BENJAMIN SCHWARTZ
1945	ASA C. CHANDLER	DONALD L. AUGUSTINE
1946	NORMAN R. STOLL	HAROLD KIRBY, JR.

Secretary-Treasurer

WILLIAM W. CORT	1925; 1926; 1927; 1928; 1929
NORMAN R. STOLL	1930; 1931; 1932

Secretary

HORACE W. STUNKARD	1933-34; 1935-36; 1937
OLIVER R. MCCOY	1938-39; 1940-41; 1942
JAMES T. CULBERTSON	1942-43; 1944-45; 1946-47

Treasurer

AUSTIN ANDREWS	1933-34; 1935-36
GILBERT F. OTTO	1937-38; 1939-40; 1944
L. E. ROZEBOOM	1941-42; 1943-44
ROBERT M. STABLER	1945-46; 1947

Council Members at Large

PAUL BARTSCH	1925-28	W. B. HERMS	1930-33
FRED C. BISHOPP	1925-28; 1929-30	BENJAMIN SCHWARTZ	1930-33
ROBERT HEGNER*	1925-27	L. R. CLEVELAND	1931
CHARLES A. KOFOID	1925	W. W. CORT	1931-34; 1935-38
B. H. RANSOM*	1925	H. E. EWING	1931-32
WILLIAM A. RILEY	1925-26; 1928-30	ERNEST C. FAUST	1931-34; 1938-41
CHARLES W. STILES*	1925; 1929-32	JOHN F. KESSEL	1932-35
ERNEST E. TYZZER	1925-26	D. H. WENRICH	1932-35; 1936
MAURICE C. HALL*	1926-29	H. E. MELENEY	1933-36
WILSON G. SMILLIE	1926-27	NORMAN R. STOLL	1933-36; 1937-40; 41
HENRY B. WARD*	1926-29	ELOISE B. CRAM	1934-37
FRANKLIN D. BARKER*	1927-30	WILBUR A. SAWYER	1934-37
J. H. ST. JOHN*	1927-28	JAMES E. ACKERT	1935-38
W. H. TALIAFERRO	1928-31	EARL C. O'ROKE	1936-39
ASA C. CHANDLER	1929-30; 1936-39	JUSTIN ANDREWS	1937-40
HARLEY J. VAN CLEAVE	1938-41	WILLARD H. WRIGHT	1942-45; 1946
ELERY R. BECKER	1939-43	HAROLD W. BROWN	1944-47
EMMETT W. PRICE	1939-43; 1944-46; 1947	HAROLD W. MANTER	1944-46
CLAY G. HUFF	1940-43; 1944-46	G. ROBERT COATNEY	1945
HORACE W. STUNKARD	1940-43	T. W. M. CAMERON	1946
DONALD L. AUGUSTINE	1941-44	MARTIN D. YOUNG	1947
RAYMOND M. CABLE	1942-45	CORNELIUS B. PHILIP	1947
GILBERT F. OTTO	1942-44; 1945		

Editorial Committee of the Journal of Parasitology

WILLIAM W. CORT, <i>Chairman</i>	1932-37	NORMAN R. STOLL, <i>Chairman</i>	1938-42; 1943
ROBERT HEGNER*	1932-34	WILLIAM A. RILEY	1934-37; 1938-42; 1943; 1944
FRANCIS M. ROOT*	1932-34	WILLIAM H. TALIAFERRO	1934-37; 1938-42; 1943

* Deceased.

HORACE W. STUNKARD, *Chairman* 1944
DAVID H. WENRICH 1944

Editorial Board of the Journal of Parasitology

CHARLES F. CRAIG	1932-33; 1934-37	CORNELIUS B. PHILIP	1939-42
MAURICE C. HALL*	1932-33	ERNEST E. TYZZER	1939-42
HENRY B. WARD*	1932-33	HAROLD W. BROWN	1940-43
ASA C. CHANDLER	1932-34; 1935-38; 1939-42	HAROLD W. MANTER	1940-43
CHARLES A. KOFOID*	1932-34; 1935-38	REGINALD D. MANWELL	1940-43
WILLIAM A. RILEY	1932-34	RICHARD P. HALL	1941-44
W. H. TALIAFERRO	1932-34	E. HAROLD HINMAN	1941-44
JAMES E. ACKERT	1932-35	JUSTUS F. MUELLER	1941-44
RICHARD P. STRONG	1932-35; 1936-39	HAROLD KIRBY	1942-45
FRED C. BISHOPP	1932-36	BENJAMIN G. CHITWOOD	1943-46
GEORGE P. LARUE	1932-36	PINCUS P. LEVINE	1943-46
DAVID H. WENRICH	1932-36; 1938-41	RUDOLF GLASER	1943-46
ERNEST C. FAUST	1933-37	LOWELL T. COGGESHALL	1944
BENJAMIN SCHWARTZ	1933-37; 1938-41; 1942-45	JOHN T. LUCKER	1944
ELERY R. BECKER	1934-35; 1936-39	NORMAN R. STOLL	1944
ROBERT MATHESON	1935-38	WILLIAM L. JELLISON	1945
OLIVER R. MCCOY	1936-39	CHARLES W. REES	1945
HENRY E. EWING	1937-40	LLOYD A. SPINDLER	1945
JOHN F. KESSEL	1937-40	RAYMOND M. CARLE	1946
HARLEY J. VAN CLEAVE	1937-40	LLOYD E. ROZEBOOM	1946
WILLIAM W. CORT	1938-41; 1942-45; 1946	RICHARD J. PORTER	1947
		WILLIAM TRAGER	1947
		ARTHUR C. WALTON	1947

List of Meeting Places

1925	Kansas City	1933	Boston	1940	Philadelphia
1926	Philadelphia	1934	Pittsburgh	1941	Dallas
1927	Nashville	1935	St. Louis	1942	(New York, cancelled)
1928	New York	1936	Atlantic City	1943	(No meeting)
1929	Des Moines	1937	Indianapolis	1944	Cleveland
1930	Cleveland	1938	Richmond	1945	St. Louis
1931	New Orleans	1939	Columbus	1946	Boston
1932	Atlantic City				

* Deceased.

IN MEMORIAM

RUDOLF W. GLASER

SEYMOUR HADWEN*

CHARLES A. KOFOID*†

SAMUEL MORRIS

HARRY PLOTZ

AMERICAN SOCIETY OF PARASITOLOGISTS

LIST OF MEMBERS¹*Honorary Foreign Members*

- BACIGALUPO, JUAN, Facultad de Medicina, Buenos Aires, Argentina.
 BAYLIS, HARRY A., British Museum (Natural History), Cromwell Road, London, S.W. 7, England.
 BRUMPT, ÉMILE, Laboratoire de Parasitologie, Faculté de Médecine, 15, Rue de l'École de Médecine, Paris VI, France.
 MARTINI, ERIC, Institut für Schiffs-und Tropenkrankheiten, Hamburg, Germany.
 SERGENT, EDMOND, L'Institut Pasteur d'Algérie, Algérie, North Africa.
 SKRJABIN, KONSTANTIN I., Institut d'Helminthologie de l'École Vétérinaire, University of Moscow, Russia.
 SWELLENGREBEL, N. H., Institute for Tropical Hygiene, Mauritskade 57, Amsterdam, The Netherlands.
 TRAVASSOS, LAURO PEREIRA, Instituto Oswaldo Cruz, Rio de Janeiro, Brazil.
 WENYON, CHARLES MORLEY, Wellcome Research Institution, 183 Euston Road, London, England.

Active Members

One life member, indicated by †

Charter members indicated by *

- *ACKERT, JAMES E., Department of Zoology, Kansas State College of Agriculture and Applied Science, Manhattan, Kansas.
 ADAMS, ALFRED J., New York State Agricultural Experiment Station, College Road, Poughkeepsie, New York.
 ALICATA, JOSEPH E., Hawaiian Agricultural Experiment Station, Honolulu, Territory of Hawaii.
 ALLEN, P. EVELYN, Department of Parasitology, Johns Hopkins University, Baltimore 5, Maryland.
 ALLEN, REX W., Zoological Laboratory, 908 Custom House, Chicago 7, Illinois.
 ALLEN, ROBERT M., Department of Pathology and Bacteriology, University of Nebraska Medical School, Omaha, Nebraska.
 ALLISON, LEONARD NEWTON, State Fish Hatchery, Grayling, Michigan.
 ALVES MEIRA, JOAO, Rua Atlantica 481, Sao Paulo, Brazil.
 AMEEL, DONALD J., Department of Zoology, Kansas State College, Manhattan, Kansas.
 ANDERSON, CYRUS V., 609 Center Street, Winona, Minnesota.
 ANDERSON, DORCAS JEAN, Department of Biology, Kent State University, Kent, Ohio.
 ANDERSON, MARLOWE G., New Mexico State College of Agriculture and Mechanical Arts, State College, New Mexico.
 ANDREWS, JOHN S., Coastal Plain Experiment Station, Tifton, Georgia.
 *ANDREWS, JUSTIN M., U. S. Public Health Service, 605 Volunteer Building, Atlanta, Georgia.
 ANNEREUX, RALPH F., Animal Pathology Laboratory, Department of Agriculture, Sacramento 4, California.
 APPLETON, SYRIL, 575 West End Avenue, New York 24, N. Y.
 ARANT, FRANK S., Zoology and Entomology Department, Alabama Polytechnic Institute, Auburn, Alabama.
 ATCHLEY, FLOYD O., Department of Biology, Defiance College, Defiance, Ohio.
 *AUGUSTINE, DONALD L., Department of Comparative Pathology, Harvard Medical School, Boston, Massachusetts.
 AUGUSTSON, S. F., California Biological Service, 1612 W. Glenoaks Boulevard, Glendale 1, California.
 AVERY, JOHN L., 1611 Monroe Street, N.W., Washington 10, D. C.
 BACHMAN, GEORGE W., Brookings Institution, 722 Jackson Place, N.W., Washington, D. C.
 BAKER, DONALD W., Parasitological Laboratory, New York State Veterinary College, Cornell University, New York.
 BAKER, EDWARD WILLIAM, U. S. Bureau of Entomology, U. S. National Museum, Washington, D. C.
 BAKER, ROLLIN H., Museum of Natural History, University of Kansas, Lawrence, Kansas.

¹ As of November 1, 1947. The last preceding list of members was published in the Journal of Parasitology, 1942, 28 Supplement: 41. Earlier lists appeared in the Journal of Parasitology in 1926, 12: 170 and in 1934, 20: 345.

- BALAMUTH, WILLIAM, Department of Zoology, Northwestern University, Evanston, Illinois.
- *BALL, GORDON H., 405 Hilgard Avenue, University of California, Los Angeles, California.
- *BANGHAM, RALPH V., Southern Fisheries Area Headquarters, Route 3, Madison, Wisconsin.
- BAROODY, BAHIJ J., Timmonsville, South Carolina.
- BARRETT, JOHN P., Chemical Research and Development Department, Armour and Company Laboratories, 1425 W. 42nd Street, Chicago 9, Illinois.
- *BARTSCH, PAUL, U. S. National Museum, Washington, D. C.
- BASNUEVE Y ARTILES, JOSE G., Apartado Correos 670, Havana, Cuba.
- *BEAUDETTE, FREDERICK R., Agricultural Experimental Station, New Brunswick, New Jersey.
- BEAVER, PAUL C., Tulane University, New Orleans, Louisiana.
- *BECKER, ELLERY R., 413 Lynn Street, Ames, Iowa.
- BELKIN, JOHN N., Biology Department, Mohawk College, Utica 5, New York.
- BELL, SAMUEL D., JR., 131 E. 74th Street, New York 21, New York.
- BELTRAN, ENRIQUE, Department of Protozoology, Institute of Public Health and Tropical Diseases, Mexico City, Mexico.
- *BENBROOK, EDWARD A., Division of Veterinary Pathology, Iowa State College, Ames, Iowa.
- BENNETT, HARRY JACKSON, 4870 Tulane Drive, Baton Rouge 14, Louisiana.
- BENNINGTON, ELWIN E., Box 41, Kelley, Iowa.
- BERRY, ELMER G., Zoology Laboratory, National Institute of Health, Bethesda, Maryland.
- *BISHOPP, FRED C., U. S. Bureau of Entomology, Washington, D. C.
- BISSINGER, LESTER L., 212 Walnut Street, S.E., Minneapolis 14, Minnesota.
- BLACK, JAMES J., Poultry Laboratory, % Landis & Brewster, Vineland, New Jersey.
- BLACKBURN, CLYDE CARLTON, % Hamilton Hall, Keene, Texas.
- BOARDMAN, EDWARD T., Rochester Museum of Arts and Sciences, 657 East Avenue, Rochester 7, New York.
- BONILLA-NAAR, ALFONSO, Carrera 4A 14-61, Bogota, Columbia.
- BOUGHTON, DONALD C., Grasselli Chemicals Department, E. I. du Pont de Nemours Company, Wilmington, Delaware.
- BOYD, ELIZABETH M., Zoology Department, Mt. Holyoke College, South Hadley, Massachusetts.
- BRACKETT, STERLING, American Cyanamid Company, Stanford Research Laboratories, Stanford, Connecticut.
- BRADBURY, ORA C., Department of Biology, Wake Forest College, Wake Forest, North Carolina.
- BRADFORD, MARY JANE, Department of Zoology, University of Wisconsin, Madison 6, Wisconsin.
- BRADIN, JOHN J., JR., Department of Parasitology, Johns Hopkins University, Baltimore, Maryland.
- BRANCH, HAZEL E., Department of Zoology, University of Wichita, Wichita, Kansas.
- BRAVO HOLLIS, MARGARITA, Patzcuaro 165, Lomas de Chapultepec, Mexico City, Mexico.
- BRENNAN, JAMES MARKS, Rocky Mountain Laboratory, Hamilton, Montana.
- BRITT, HENRY GRADY, Box 192, Colerain, North Carolina.
- BROOKE, MARION M., U. S. Public Health Service, 291 Peachtree Street, Atlanta, Georgia.
- BROOKMAN, BERNARD, U. S. Public Health Service, % Hooper Foundation, University of California, San Francisco, California.
- *BROOKS, FRANK G., Cornell College, Mount Vernon, Iowa.
- BROOKS, THOMAS JOSEPH, JR., Department of Parasitology, University of Mississippi, University, Mississippi.
- *BROWN, HAROLD W., College of Physicians and Surgeons, Delamar Institute of Public Health, Columbia University, 600 W. 168th Street, New York, New York.
- BROWNE, PATRICK, 250 Bellevue Avenue, Trenton, New Jersey.
- BURLESON, GRETCHEN L., Department of Zoology, University of California at Los Angeles, Los Angeles, California.
- BURROWS, ROBERT, P. O. Box 189, Columbia, South Carolina.
- BUTTS, DONALD C. A., United Fruit Company, Medical Division, Puerto Limon, Costa Rica, Central America.
- BYRD, ELON E., Department of Zoology, University of Georgia, Athens, Georgia.
- CABALLERO Y CABALLERO, EDUARDO, Instituto de Biologia, Chapultepec Casa del Lego, Mexico City, Mexico.
- CABLE, RAYMOND, Department of Biology, Purdue University, W. Lafayette, Indiana.
- CAMERON, JOHN A., Baylor University, College of Dentistry, College and Gaston Avenues, Dallas, Texas.
- CAMERON, THOMAS W. M., MacDonald College, Quebec, Canada.
- CAMMACK, MARGARET ROSALYN, Department of Bacteriology, University of Texas, Medical Branch, Galveston, Texas.
- CANTRELL, WILLIAM F., 1591 South Lumpkin Street, Athens, Georgia.
- *CARROLL, MITCHELL, Department of Biology, Franklin and Marshall College, Lancaster, Pennsylvania.

- CARSON, HAMPTON LAWRENCE, Department of Zoology, Washington University, St. Louis, 5, Missouri.
- CARTER, FRANKLIN, 2 Brattle Circle, Cambridge, Massachusetts.
- CASE, ARTHUR A., Department of Veterinary Science, Ohio State University, Columbus, Ohio.
- CASIS-SACRE, GUILLERMO, School of Medicine, National University of Mexico, Mexico City, Mexico.
- CAUSEY, OTIS R., Caixa Postal 49, Rio de Janeiro, Brazil.
- CAUTHEN, GEORGE E., Regional Animal Disease Research Laboratory, U. S. Bureau of Animal Industry, P. O. Drawer 952, Auburn, Alabama.
- CHAMBERLAIN, ROY WILLIAM, Department of Parasitology, Johns Hopkins University, Baltimore 5, Maryland.
- *CHANDLER, ASA C., Department of Biology, Rice Institute, Houston, Texas.
- CHAPMAN, J. W., Walterboro, South Carolina.
- CHAUHAN, B. S., Zoological Survey of India, Kaiser Castle, Benares, India.
- CHAVEZ-GARCIA, CARLOS E., Faculty of Veterinary Medicine, Las Palmas, Barranca, Lima, Peru.
- CHEATUM, E. L., Bureau of Wildlife Investigation, New York State Conservation Department, Delmar, New York.
- CHEN-SUI-FONG, 2838 South Abingdon Street, Fairlington, Arlington, Virginia.
- CHERNOFF, HARRY A., 584 Linwood Avenue, Buffalo 9, New York.
- CHIANG, TZE SHENG, 2621 Knapp Street, Ames, Iowa.
- CHINO, TA-HSIUNG, Department of Tropical Medicine, New Orleans, Louisiana.
- *CHRISTIE, JESSIE R., Division of Nematology, Plant Industry Station, Beltsville, Maryland.
- CHURCHILL, HELEN, Hollins College, Virginia.
- *CLARK, HERBERT C., Gorgas Memorial Laboratory, Apartado 1252, Panama, Republic de Panama.
- COATNEY, ROBERT G., Division of Physiology, National Institute of Health, Bethesda, Maryland.
- COFFIN, DAVID L., 180 Longwood Avenue, Boston 15, Massachusetts.
- COGGESHALL, LOWELL T., Department of Medicine, University of Chicago, Chicago, Illinois.
- CONNELL, FRANK H., Dartmouth College, Hanover, New Hampshire.
- CORIA, NICHOLAS A., Rockefeller Institute for Medical Research, Princeton, New Jersey.
- CORPSON, RUTH ALEXANDRA, Department of Pathology, Charity Hospital, New Orleans, Louisiana.
- *CORT, WILLIAM W., Department of Parasitology, Johns Hopkins University, Baltimore 5, Maryland.
- COULSTON, FREDERICK, 1604 North Franklin Street, Wilmington, Delaware.
- *COVENTRY, FRANCES A., 158 Wilson Drive, Lancaster, Pennsylvania.
- †*CRAIG, CHARLES F., 239 West Lullwood Avenue, San Antonio, Texas.
- *CRAM, ELOISE B., Zoology Laboratory, National Institute of Health, Bethesda 14, Maryland.
- CRAWFORD, WILEY W., 1305 Parrett Street, Evansville 13, Indiana.
- CROSS, JOY BARNES, Department of Preventive Medicine, University of Texas Medical School, Galveston, Texas.
- CROWELL, ROBERT MERRILL, Department of Biology, Bowling Green State University, Bowling Green, Ohio.
- CUCKLER, ASHTON CLINTON, Merck Institute for Therapeutic Research, Rahway, New Jersey.
- CULBERTSON, JAMES T., Department of Bacteriology and Parasitology, University of Arkansas, Little Rock, Arkansas.
- DALMAT, HERBERT T., 247 Audubon Avenue, New York 33, New York.
- DANIEL, GEORGE E., National Cancer Institute, Bethesda 14, Maryland.
- D'ANTONI, JOSEPH S., 1825 Calboun Street, New Orleans 15, Louisiana.
- DAVIS, GORDON E., Rocky Mountain Laboratory, Hamilton, Montana.
- DAVIS, HELEN EDITH, Department of Zoological Sciences, University of Oklahoma, Norman, Oklahoma.
- DAVIS, LEONARD R., Regional Animal Disease Research Laboratory, U. S. Bureau of Animal Industry, Auburn, Alabama.
- DEGIUSTI, DOMINIC L., Department of Biology, Catholic University of America, Washington, D. C.
- DEANE, LEONIDAS M., Benjamin Constant 724, Belem Para, Brazil.
- DEHNOSTEL, NELLIE G., 178 Iddings Avenue, N.E., Warren, Ohio.
- DELAUNE, ELAINE T., Box 3482, University, Louisiana.
- DENTON, FRED J., Department of Bacteriology and Public Health, Georgia Medical School, Augusta, Georgia.
- DERBYSHIRE, RUSSELL C., University of Omaha, 60th and Dodge Street, Omaha 1, Nebraska.
- DETURK, WILLIAM E., Department of Pharmacology, Vanderbilt University, School of Medicine, Nashville 4, Tennessee.

- DEVOLT, HAROLD MOON, University of Maryland, Department of Pathology, College Park, Maryland.
- DIAMOND, LOUIS STANLEY, Department of Zoology, University of Minnesota, Minneapolis 14, Minnesota.
- DICKERMAN, EUGENE E., Bowling Green State University, Bowling Green, Ohio.
- DINGEE, RUTH FOSTER, Route 1, Box 879, Maryville, Washington.
- DOBROVOLNY, CHARLES G., Zoology Department, University of New Hampshire, Durham, New Hampshire.
- DOETSCHMAN, WILLIS H., #9 Ronada Avenue, Oakland 11, California.
- DONALDSON, ALAN W., 605 Volunteer Bldg., Atlanta 3, Georgia.
- DOTTERER, JOHN E., 510 Gulf Street, Sanford, North Carolina.
- DOUGHERTY, ELLSWORTH C., Department of Medical Physics, University of California, Berkeley 4, California.
- DOUGLAS, JAMES RUSSELL, Division of Entomology and Parasitology, University of California, Davis, California.
- *DOVE, WALTER E., Defense Highway, Gambrills, Maryland.
- DOWNES, WILBUR G., 26 Colle Viena, Mexico D.F., Mexico.
- *DRAKE, CARL J., Science Building, Iowa State College, Ames, Iowa.
- DUFF, FRATIS L., Headquarters, 315-Composite Wing, APO #929, c/o Post Master, San Francisco, California.
- DUNLAP, JACK SHERWIN, Department of Bacteriology, Michigan State College, E. Lansing, Michigan.
- DUNN, MARY CATHERINE, Culver Stockton College, Canton, Missouri.
- EADS, RICHARD B., Box 1305, College Station, Texas.
- EARLE, HILTON H., JR., 712 E. Carocas Street, Tampa, Florida.
- EDELMAN, MORTON HENRY, 2 East 77th Street, New York City New York.
- EDGAR, ALLEN S., Poultry Department, Alabama Polytechnical Institute, Auburn, Alabama.
- EISENBRANDT, LESLIE L., Department of Biology, University of Kansas City, Kansas City, Missouri.
- ELGINDY, MOHAMMED S., 611 Church Street, Ann Arbor, Michigan.
- ELISHEWITZ, HAROLD, Munoz A Pedrera 40, Caracas, Venezuela.
- EVERITT, MARTHA GRACE, Department of Tropical Medicine, Tulane University, New Orleans, Louisiana.
- *EWING, HENRY E., U. S. National Museum, Washington, D. C.
- FALLIS, ARTHUR M., Ontario Research Foundation, 43 Queen's Park, Toronto 5, Canada.
- FARNER, DONALD SANKEY, Department of Zoology, State College of Washington, Pullman, Washington.
- FARR, MARION M., Zoological Division, U. S. Bureau of Animal Industry, Beltsville, Maryland.
- FAUST, ERNEST C., Department of Tropical Medicine, Tulane University, New Orleans, Louisiana.
- FELSENFELD, OSCAR, P. O. Box 32, San Juan 1, Puerto Rico.
- FERGUSON, MALCOLM S., Rockefeller Institute for Medical Research, Princeton, New Jersey.
- FERRIS, DEAM H., 717 E. 13th St., Des Moines, Iowa.
- FIGGATT, WILLIAM BURNETT, Division of Zoology, National Institute of Health, Bethesda 14, Maryland.
- FILES, VIRGINIA SHAULIS, Zoological Laboratory, National Institute of Health, Bethesda 14, Maryland.
- FINKELSTEIN, SAMUEL M., 825 Blue Hill Avenue, Dorchester, Massachusetts.
- FISCHTHAL, JACOB H., Biology Division Laboratory, Wisconsin Conservation Department, Spooner, Wisconsin.
- FISHER, WILTON MONROE, Department of Public Health and Preventive Medicine, Baylor University, College of Medicine, Houston, Texas.
- FOLSE, DEAN SYDNEY, Zoology Department, University of Minnesota, Minneapolis 14, Minnesota.
- FORBES, WILLIAM C., 190 Main Street, Bridgton, Maine.
- FOSTER, AUREL O., Zoological Division, U. S. Bureau of Animal Industry, Beltsville, Maryland.
- FOWLER, JAMES ABBOTT, The Sidwell Friends School, 3901 Wisconsin Avenue, Washington 16, D. C.
- FOX, IRVING, Department of Medical Zoology, School of Tropical Medicine, San Juan, Puerto Rico.
- FREEMAN, REINO SAMUEL, Department of Entomology, University Farm, St. Paul 1, Minnesota.
- FREITAS, MOACYR G., Escola Superior de Veterinaria, Caixa Postal 567, Belo Horizonte, Minas Gerais, Brazil.
- FRENKEL, JACOB, Division of Pathology, Medical School, San Francisco 22, California.

- FRICK, LYMAN P., Department of Parasitology, Army Medical Center, Washington 12, D. C.
- FULLER, HENRY SHEPARD, Department of Preventive Medicine, Bowman Gray School of Medicine, Winston-Salem 7, North Carolina.
- FURMAN, DEANE P., 197 Amherst Avenue, Berkeley 8, California.
- GALLO, PIERO, Apartado de Cooov No. 1308, Caracas, Venezuela.
- GEIMAN, QUENTIN M., Department of Comparative Pathology, Harvard Medical School, Boston, Massachusetts.
- GERMAN, WILLIAM McKEE, Department of Pathology, University of Cincinnati, Cincinnati, Ohio.
- GIBSON, COLVIN L., 3103 Oakwood, Route 7, Ann Arbor, Michigan.
- GIFFEN, HORACE KERR, Department of Pathology, Western Reserve University, Cleveland, Ohio.
- GILCHRIST, HAZEL B., School of Public Health, University of North Carolina, Chapel Hill, North Carolina.
- GINGRICH, WENDELL, Department of Bacteriology, University of Texas Medical School, Galveston, Texas.
- GOBLE, FRANS C., Sterling-Winthrop Research Institute, Rensselaer, New York.
- GOLDHAFT, TEUIS M., Box 248, Vineland, New Jersey.
- GOOD, WILLIAM MURRAY, 219 Columbia Blvd., Waterbury, Connecticut.
- GOODCHILD, CHAUNCEY G., Department of Biology, State Teachers College, Springfield, Missouri.
- GOODMAN, JOHN D., Department of Zoology, University of Michigan, Ann Arbor, Michigan.
- GOODNIGHT, CLARENCE J., Department of Biology, Biology Annex, Purdue University, W. Lafayette, Indiana.
- GOODWIN, MELVIN H., JR., 605 Volunteer Building, Atlanta, Georgia.
- GORDON, WILLIAM HENRY, 1102 David Whitney Building, Detroit, Michigan.
- GOULDING, ROBERT L., Department of Zoology and Entomology, Ohio State University, Columbus, Ohio.
- GRAHAM, GEORGE L., Rockefeller Institute for Medical Research, Princeton, New Jersey.
- GRAVATT, MARGARET SPENCER, School of Public Health, University of North Carolina, Chapel Hill, North Carolina.
- GREEN, NANCY K. V., 252 Roslyn Road, Winnepeg, Manitoba, Canada.
- GRIFFIN, ANGUS McIVOR, Department of Bacteriology, George Washington University, Medical School, Washington 5, D. C.
- GROCOTT, ROBERT GREENWOOD, Board of Health Laboratory, Ancon, Canal Zone.
- GROSCH, DANIEL SWARTWOOD, Zoology Department, North Carolina State College, Raleigh, North Carolina.
- GUSTAFSON, PAUL V., 4217 E. 22nd Avenue, Spokane 10, Washington.
- *HALL, RICHARD P., Department of Biology, New York University, University Heights, New York, New York.
- HALSEY, HARVEY RANDOLPH, Department of Zoology, College of Pharmacy, Columbia University, New York, New York.
- HALSTEAD, BRUCE W., Department of Medical Zoology, School of Tropical and Preventive Medicine, Loma Linda, California.
- HAMANN, CECIL B., Asbury College, Wilmore, Kentucky.
- HAMMOND, DATUS M., Department of Zoology, Utah State Agricultural College, Logan, Utah.
- *HANNUM, CLAIR A., Department of Zoology, University of Wichita, Wichita, Kansas.
- HANSEN, MERLE F., Department of Zoology, University of Nebraska, Lincoln, Nebraska.
- HARDCASTLE, A. BASCOM, 4330 Clogett Road, Hyattsville, Maryland.
- HARKEMA, REINARD, Department of Zoology, State College, Raleigh, North Carolina.
- HART, THOMAS A., % American Consulate, Cochabamba, Bolivia, South America.
- *HARTMAN, ERNEST, Chicago Medical School, 710 South Wolcott Avenue, Chicago 12, Illinois.
- HARWOOD, PAUL D., 337 W. Walnut Street, Ashland, Ohio.
- HASSON, ELEANOR V., 3807 Nicholson Street, Hyattsville, Maryland.
- *HATHAWAY, EDWARD S., Department of Zoology, Tulane University, New Orleans, Louisiana.
- HAUSCHKA, THEODORE S., The Lankenau Hospital, Girard and Corinthian Avenue, Philadelphia 30, Pennsylvania.
- HAWKINS, PHILIP A., Department of Bacteriology, Michigan State College, East Lansing, Michigan.
- HAZARD, FRANK O., Department of Zoology, Wilmington College, Wilmington, Ohio.
- HEADLEE, WILLIAM H., Division of Clinical Pathology, Indiana University School of Medicine, Indianapolis, Indiana.
- HEDRICK, LESLIE, Department of Biology, Illinois Institute of Technology, Chicago 16, Illinois.
- HENDRICKS, JAMES R., School of Public Health, University of North Carolina, Chapel Hill, North Carolina.
- HERBER, ELMER C., 416 W. South Street, Carlisle, Pennsylvania.

- HERMAN, CARLTON M., California Division of Fish and Game, Ferry Building, San Francisco, California (11).
- *HERMS, WILLIAM B., Agricultural Hall, University of California, Berkeley 4, California.
- *HERRICK, CHESTER A., University of Wisconsin, Department of Zoology, Madison, Wisconsin.
- HERSHBERGER, LLOYD R., National Institute of Health, Bethesda 14, Maryland.
- HERTIG, MARSHALL, Headquarters PCD., APO 834, c/o Post Master, New Orleans, Louisiana.
- *HETHERINGTON, DUNCAN C., Duke Hospital, Durham, North Carolina.
- HIGHBY, PAUL RICHARD, 2082 Como Avenue, West, Saint Paul, Minnesota.
- HILL, CHARLES H., Zoological Division, U. S. Bureau of Animal Industry, Beltsville, Maryland.
- HILL, HOWARD R., 4262 Angeles Vista Boulevard, Los Angeles, California.
- HINMAN, HAROLD E., Health and Safety Department, Tennessee Valley Authority, Wilson Dam, Alabama.
- HITCHCOCK, DOROTHY JEAN, Research Laboratory, Ciba Pharmaceutical Products Inc., Summit, New Jersey.
- HOBMAIER, M., 1505 Masonic Avenue, San Francisco, California.
- HOFF, CLAYTON C., Department of Biology, University of New Mexico, Albuquerque, New Mexico.
- HOFFMAN, GLENN LYLE, Quadrangle Cottage 25, Iowa City, Iowa.
- HOGUE, MARY J., Department of Anatomy, University of Pennsylvania, Philadelphia, Pennsylvania.
- HOISINGBERG, BRONISLAW, Department of Zoology, University of California, Berkeley, California.
- HOK, KAROL ANTON, Department of Zoology, University of California, Berkeley, California.
- *HOLL, FRED J., Department of Biology, University of Buffalo, Buffalo, New York.
- HOLLIS, AUDREY MARION, Department of Tropical Medicine, Tulane University, New Orleans, Louisiana.
- HOOD, MARION WINIFRED, Department of Pathology, Charity Hospital, New Orleans 13, Louisiana.
- HOOGHKIRK, EDWARD O., Department of Zoology, University of New Hampshire, Durham, New Hampshire.
- HOPLA, CLUFF E., 360 East Eighth South Street, Springville, Utah.
- HOPP, WILLIAM B., Department of Biology, Purdue University, Lafayette, Indiana.
- HORSFALL, MARGERY W., Route 2, Fairport, New York.
- HSIUNG, T. S., College of Agriculture, National University of Peking, West Suburb, Peiping, China.
- HUDSON, CHARLES B., Agriculture Experiment Station, New Brunswick, New Jersey.
- *HUFF, CLAY G., Naval Medical Research Institute, Bethesda, Maryland.
- HUGHES, CHESTER R., 511 W. 5th Avenue, Stillwater, Oklahoma.
- HUMES, ARTHUR G., Department of Biology, Boston University, Boston, Massachusetts.
- HUNNINEN, ARNE V., 1303 Electric Boulevard, Alliance, Ohio.
- HUNT, OLEN E., U. S. Bureau of Entomology and Plant Quarantine, 7300 Wingate, Room 206, Houston, Texas.
- HUNT, STEGER J., Zoology Department, University of Michigan, Ann Arbor, Michigan.
- *HUNTER, GEORGE W., III, Department of Parasitology, 406 Medical General Laboratory, APO #500, c/o Post Master, San Francisco, California.
- HUNTER, WANDA S., Department of Zoology, Duke University, Durham, North Carolina.
- HURLBUT, HERBERT S., Naval Medical Research Institute, Bethesda 14, Maryland.
- HUSSEY, KATHLEEN L., Delamar Institute, Columbia University, New York, New York.
- INGALLS, MABEL S., Saltsbury Mills, New York.
- JACHOWSKI, LEO, 76 Prospect Street, Kensington, Maryland.
- JACOBS, LEON, Zoology Laboratory, National Institute of Health, Bethesda, Maryland.
- JAMESON, EVERETT WILLIAMS, JR., Department of Zoology, Cornell University, Ithaca, New York.
- JANKIEWICZ, HARRY A., 1320 E. Windsor Avenue, Glendale, California.
- JARCHO, SAUL, Mt. Sinai Hospital, Fifth Avenue and 100th Street, New York 29, New York.
- JEFFREY, GEOFFREY MARRON, Department of Zoology, University of Bridgeport, Bridgeport, Connecticut.
- JELLISON, WILLIAM L., Rocky Mountain Laboratory, U. S. Public Health Service, Hamilton, Montana.
- JEWELL, ROSS LYMAN, School of Veterinary Medicine, Kansas State Agricultural College, Manhattan, Kansas.
- JOHNSTONE, HERBERT G., Department of Medical Parasitology and Mycology, University of California, Medical School, Medical Center, San Francisco 22, California.
- JONES, ARTHUR W., Department of Zoology and Entomology, Agricultural Station, Knoxville 16, Tennessee.

- JONES, ELMER ALSBROOK, Division of Parasitology, Army Medical School, Washington, D. C.
JONES, MYRNA F., Division of Zoology, National Institute of Health, Bethesda 14, Maryland.
KAGAN, IRVING GEORGE, Department of Zoology, University of Michigan, Ann Arbor, Michigan.
KAHN, MORTON C., Department of Preventive Medicine and Public Health, Cornell University Medical College, New York, New York.
KALANTAR, LEVON, Department of Biology, New York University, New York 53, New York.
KATES, KENNETH C., Zoological Division, U. S. Bureau of Animal Industry, Beltsville, Maryland.
KAUFMAN, BERNARD, 2442 Broadway, San Francisco, California.
KEEGAN, HUGH LAWRENCE, 7th Service Command, Medical Laboratory, Ft. Omaha 11, Nebraska.
KEMPER, HARRY ELVIN, Zoological Division, U. S. Bureau of Animal Industry, P. O. Box 464, Albuquerque, New Mexico.
KERR, JOHN A., Rockefeller Foundation, Viale Regina Margherita, Rome, Italy.
KERR, KATHEL B., 503 Cedar Street, Charles City, Iowa.
*KESSEL, JOHN F., School of Medicine, University of Southern California, Los Angeles, California.
KHAW, OO-KEK, % Director of Medical Service, Civic Center, P. O. Box 2058, Shanghai, China.
*KING, WILLARD V., U. S. Bureau of Entomology, BqX 3391, Orlando, Florida.
KINGSBURY, DONALD L., 20 Nelson Street, Framingham, Massachusetts.
KIRBY, HAROLD, JR., Department of Zoology, University of California, Berkeley 4, California.
KLECKNER, ALBERT L., Research Division, Pitman-Moore Company, Indianapolis 6, Indiana.
KNIGHT, ALVA ALLEN, Department of Clinical Medicine, University of Illinois, Medical School (Rush), Chicago, Illinois.
KNISKERN, VERNE B., Department of Zoology, University of Michigan, Ann Arbor, Michigan.
KOLRUSS, FRED J., University of Portland, Portland, Oregon.
KOHLS, GLEN M., U. S. Public Health Service, Rocky Mountain Laboratory, Hamilton, Montana.
KOONZ, CARL H., Chemical Laboratory, Swift and Company, Union Stock Yards, Chicago, Illinois.
KOURI, PEDRO, 5a Avenida y Colle 78, Reparto Playa Miramar, Marianao, Havana, Cuba.
KRUIDENIER, FRANCIS, 812 E. Catherine Street, Ann Arbor, Michigan.
KRULL, WENDELL H., Division of Veterinary Medicine, Colorado State College, Fort Collins, Colorado.
*KUDO, RICHARD R., Department of Zoology, University of Illinois, Urbana, Illinois.
KUITUMEN, ELLA, Department of Epidemiology, School of Hygiene, University of Toronto, Toronto, Canada.
KUNTZ, ROBERT E., Naval Medical Research Institute, Bethesda 14, Maryland.
*LAAKE, ERNEST W., Box 232, Kerrville, Texas.
LAIRD, RAYMOND L., Division of Preventive Medicine, University of Tennessee, 874 Union Avenue, Memphis 3, Tennessee.
LARSH, JOHN E., JR., School of Public Health, University of North Carolina, Chapel Hill, North Carolina.
*LARSON, MARY E., Department of Zoology, University of Kansas, Lawrence, Kansas.
*LARUE, GEORGE R., Department of Zoology, University of Michigan, Ann Arbor, Michigan.
LAUGHLIN, ELMER HENRY, Long Island Medical College, 281 Henry Street, Brooklyn 2, New York.
LAUZUN, VIRGINIA, Box 67, Route #2, Lansing, Michigan.
LEADINGHAM, ROY S., 3 Park Lane, N.E., Atlanta, Georgia.
LEVI CASTILLO, ROBERTO, P. O. Box #759, Guayaquil, Ecuador, South America.
LEVIN, ARTHUR J., 660 Virginia Avenue, Atlanta, Georgia.
LEVINE, NORMAN DION, Department of Veterinary Parasitology, College of Veterinary Medicine, University of Illinois, Urbana, Illinois.
LEVINE, PINCUS P., New York State Veterinary College, Cornell University, Ithaca, New York.
LIEBERTHAL, MILTON M., Professional Building, Bridgeport, Connecticut.
LILLICK, LOIS C., Department of Bacteriology, New York Medical College, 105th Street and Fifth Avenue, New York 29, New York.
LILLIGREN, BETTY LOU, Research Division, Parke Davis and Company, Detroit, Michigan.
LINCICOME, DAVID R., Department of Medical Microbiology, School of Medicine, University of Wisconsin, Madison, Wisconsin.
LINDQUIST, WILLIAM D., Department of Parasitology, Johns Hopkins University, Baltimore 5, Maryland.
LIPOVSKY, LOUIS J., Department of Entomology, Kansas University, Lawrence, Kansas.
LOEWEN, SOLOMON L., Tabor College, Hillsboro, Kansas.
LUCKER, JOHN T., Zoological Division, U. S. Bureau of Animal Industry, Beltsville, Maryland.
LUDWIG, FRANCIS W., Monastery, Villanova, Pennsylvania.
LUND, EVERETT E., Disease Research Unit, U. S. Rabbit Experiment Station, Fontana, California.
LUNGSTROM, LEON GEORGE, 206 South Cedar Street, Lindsborg, Kansas.

- LUTTERMOSER, GEORGE W., Zoology Laboratory, National Institute of Health, Bethesda 14, Maryland.
- *LYNCH, JAMES E., University of Washington, Seattle 5, Washington.
- *LYNCH, KENNETH M., Medical College of South Carolina, Charleston, South Carolina.
- LYONS, RANDOLPH, Department of Clinical Medicine, Tulane University, New Orleans, Louisiana.
- MACLULICH, DUNCAN A., 144 Mavety Street, Toronto, Canada.
- MACY, RALPH W., Department of Biology, Reed College, Portland, Oregon.
- MADALENA, ROSE L., 1105 Terrace Drive, Napa, California.
- *MAGATH, THOMAS B., Mayo Clinic, Rochester, Minnesota.
- MALDONADO, JOSE FERNANDO, Department of Medical Zoology, School of Tropical Medicine, San Juan, Puerto Rico.
- MANGRUM, JAMES F., Biology Department, Box 203, Texas Agricultural and Mechanical College Station, Texas.
- MANN, MARY ELIZABETH, Zoology Laboratory, National Institute of Health, Bethesda, Maryland.
- *MANTER, HAROLD W., Department of Zoology, University of Nebraska, Lincoln, Nebraska.
- MANWELL, REGINALD DICKINSON, Department of Zoology, Syracuse University, Syracuse, New York.
- MARGOLIES, JEROME BENARD, 853 Bushwick Avenue, Brooklyn, New York.
- MARKELL, EDWARD K., 1120 Cragmont Avenue, Berkeley 8, California.
- MARQUEZ-ESCOBEDO, MANUEL B., Calle de Puebla 305, Mexico City, Mexico.
- MARTIN, DONALD STOVER, Department of Bacteriology, Duke University Medical School, Durham, North Carolina.
- *MARTIN, H. M., School of Veterinary Medicine, University of Pennsylvania, Philadelphia, Pennsylvania.
- MARTIN, WALTER E., Zoology Department, University of Southern California, Los Angeles 7, California.
- MARTINEZ-BAEZ, MANUEL, Atoyac 53, Mexico City, Mexico.
- *MATHESON, ROBERT, Department of Entomology, Cornell University, Ithaca, New York.
- MAUSS, EVELYN ABRAMS, 443 Beach 138 Street, Belle Harbor, New York.
- MAXWELL, ELMER STEPHENS, 190 N. Upper Street, Lexington, Kentucky.
- MAXWELL, NANCY, Lankenau Hospital Research Institute, Corinthian and Girard Avenues, Philadelphia 30, Pennsylvania.
- MAYFIELD, ORLEY J., Dr. Salsbury's Laboratories, Charles City, Iowa.
- *MAYHEW, ROY L., Department of Zoology, Louisiana State University, University, Louisiana.
- MAZZOTTI, LUIS, Puebla 256, Mexico, D.F., Mexico.
- MCCOY, OLIVER R., % Rockefeller Foundation, 20 Rue de la Baume, Paris 8, France.
- MCCULLOCH, IRENE A., Allan Hancock Foundation, University of Southern California, 3551 University Avenue, Los Angeles, California.
- MCDOWELL, JOHN W., South Hall, Colorado A. and M. College, Ft. Collins, Colorado.
- McFARLANE, SAMUEL H., Aurora College, Aurora, Illinois.
- *MCINTOSH, ALLEN, Zoological Division, U. S. Bureau of Animal Industry, Beltsville, Maryland.
- McKAY, FRANCIS, Dr. Salsbury's Laboratories, Charles City, Iowa.
- McMULLEN, DONALD B., 406th Medical General Laboratory, APO #500, % Post Master, San Francisco, California.
- McNAUGHT, JAMES B., Department of Pathology, University of Colorado, School of Medicine, 4200 E. Ninth Street, Denver 7, Colorado.
- McNEIL, CHARLES W., 1800 C Street, Pullman, Washington.
- McNEIL, ETHEL C. E., Department of Veterinary Science, University of California, Davis, California.
- McQUAY, RUSSELL MICHAEL, JR., Department of Tropical Medicine, Tulane University, New Orleans, Louisiana.
- MEINKOTH, NORMAN A., Martin Biological Laboratory, Swarthmore College, Swarthmore, Pennsylvania.
- *MELENEY, HENRY E., Department of Preventive Medicine, New York University College of Medicine, 477 First Avenue, New York, New York.
- MELVIN, DOROTHY MAE, U. S. Public Health Service, 291 Peachtree Street, Atlanta, Georgia.
- MERRILL, GEORGE G., Baldwin, Maryland.
- *MESERVE, FRANK G., Department of Biology, Bowling Green State University, Bowling Green, Ohio.
- MEYER, MARVIN C., Zoology Department, University of Maine, Orono, Maine.
- MICKS, DON WILFRED, Department of Parasitology, Johns Hopkins University, Baltimore 5, Maryland.

- MILES, VIRGIL I., Office of Service Command Engineer, Hdq. 6th Service Command, A.S.F., 20 N. Wacker Drive, Chicago 6, Illinois.
- MILLER, EDWIN L., Department of Biology, Lawrence College, Appleton, Wisconsin.
- *MILLER, HARRY M., JR., Room 5500, 49 West 49th Street, New York, New York.
- MILLER, JOAN EMILY, Illinois State Department of Health, 215 South Washington Street, Taylorville, Illinois.
- MIZELLE, JOHN D., Department of Zoology, University of Notre Dame, Notre Dame, Indiana.
- MOHR, JOHN L., Department of Zoology, Allan Hitchcock Foundation, University of Southern California, University Park, Los Angeles, California.
- MÖNNIG, HERMANN H., 25 Murray Street, Brooklyn, Pretoria, South Africa.
- MOORE, DONALD H., 406 Mt. Holly Street, Baltimore 29, Maryland.
- MOORE, DONALD V., Department of Preventive Medicine, New York University, Medical School, New York, New York.
- MOREHOUSE, NEAL F., Dr. Salsbury's Laboratories, Charles City, Iowa.
- MORGAN, BANNER BILL, Department of Veterinary Science, University of Wisconsin, Madison, Wisconsin.
- MORRIS, JOSEPH ANTHONY, Patuxent Research Refuge, Bowie, Maryland.
- MORTON, WILLIAM MARKHAM, 2725 Montlake Boulevard, Seattle, Washington.
- MOSS, EMMA S., Department of Pathology, Charity Hospital, New Orleans, Louisiana.
- MOST, HARRY, 477 First Avenue, New York, New York.
- *MUELLER, JUSTUS F., Department of Bacteriology and Public Health, Syracuse University, Syracuse, New York.
- MUELLER, MILTON GOTTLIEB, 1055 Clark Avenue, Detroit 9, Michigan.
- *MUGRAGE, E. R., 4200 East 9th Avenue, Denver, Colorado.
- NEHAUL, BALBIR BALL GREEN, Central Medical Laboratory, Middle Street, Georgetown, British Guiana.
- NEGhme-RODRIGUEZ, AMADOR, Department of Parasitology, Public Health Service, Santiago, Chile.
- NELSON, CLIFFORD E., Box 143, Woods Hole, Massachusetts.
- *NELSON, THURLOW C., Department of Zoology, Rutgers University, New Brunswick, New Jersey.
- NEUMAYER, ELEANOR MAY, 910 Delaware Avenue, Wilmington, Delaware.
- NEVIN, F. REESE, RFD No. 1, Plattsburg, New York.
- NEWTON, WALTER L., U. S. Public Health Service, U. S. Public Health Service Hospital, Lexington, Kentucky.
- NIELSEN, GORDON E., Zoology Department, State University of Iowa, Iowa City, Iowa.
- NIGRELLI, ROSS F., Aquarium, New York Zoology Park, 185th Street and Southern Boulevard, Bronx, New York, New York.
- NOBLE, ELMER R., 1250 Dover Lane, Santa Barbara, California.
- *NOLAN, MABELLE O., National Institute of Health, Bethesda, Maryland.
- *NOLAND, LOWELL E., Biology Building, University of Wisconsin, Madison, Wisconsin.
- NOLF, LUTHER O., Department of Zoology, State University of Iowa, Iowa City, Iowa.
- OFFUTT, EDWARD P., JR., Department of Bacteriology, School of Medicine and Dentistry, University of Rochester, Rochester, New York.
- OHL, HARRY E., General Delivery, Angwin, California.
- OLIVER-GONZALEZ, JOSE, School of Tropical Medicine, San Juan, Puerto Rico.
- OLIVIER, LOUIS J., Zoology Laboratory, National Institute of Health, Bethesda 14, Maryland.
- OLSEN, LELAND S., Department of Zoology, University of Nebraska, Lincoln, Nebraska.
- OLSEN, WILFORD O., Zoological Division, U. S. Bureau of Animal Industry, % Texas Experiment Station, Angleton, Texas.
- *O'ROKE, EARL C., 4052 Natural Science Building, University of Michigan, Ann Arbor, Michigan.
- *OSBORN, HERBERT, Ohio State University, Columbus, Ohio.
- OSBURN, RAYMOND C., Ohio State University, Columbus, Ohio.
- OTTO, GILBERT F., Department of Parasitology, Johns Hopkins University, Baltimore 5, Maryland.
- OWEN, WILLIAM B., Department of Zoology, University of Wyoming, Laramie, Wyoming.
- PACKCHANIAN, A., University of Texas Medical School, Galveston, Texas.
- PALMER, EDDY D., Walter Reed General Hospital, Washington, D. C.
- PAPINEAU, ALBAN, Plymouth Clinic, Plymouth, North Carolina.
- *PARMAN, DANIEL C., Box 509, Uvalde, Texas.
- PAUL, ALLARD A., 330 East 18th Street, New York City, New York.
- *PAYNE, GEORGE C., Room 5500, 49 West 49th Street, New York 20, New York.
- *PEARSE, ARTHUR S., Zoology Department, Duke University, Durham, North Carolina.
- PENNER, LAWRENCE R., Department of Zoology, University of Connecticut, Storrs, Connecticut.

- PEQUEÑO, EDUARDO AGUIRRE, Apartado 897, Monterrey, N.L., Mexico.
- PEREIRA, CLEMENTE, Instituto Biológico de São Paulo, Caixa Postal 119A, São Paulo, Brazil.
- PHILIP, CORNELIUS B., Rocky Mountain Spotted Fever Laboratory, U. S. Public Health Service, Hamilton, Montana.
- PIMENTAL IMBERT, MANUEL FELIPE, University of Santo Domingo and International Hospital, Trujillo City, Dominican Republic.
- PIPKIN, ALAN COLLINS, Department of Bacteriology and Parasitology, Medical Division, American University of Beirut, Beirut, Lebanon.
- POHL, MARIAN E., Insect Control Committee, National Academy of Science, 2101 Constitution Avenue, Washington 25, D. C.
- POINDEXTER, HILDRUS A., 1430 AAF Base Unit, APO #194, Br. Unit #2, c/o Post Master, New York, New York.
- PORTER, ANNIE, Zoological Society of London, Regent's Park, London, N.W. 8, England.
- PORTER, DALE A., Regional Animal Disease Research Laboratory, U. S. Bureau Animal Industry, Auburn, Alabama.
- PORTER, RICHARD JANVIER, School of Public Health, University of Michigan, Ann Arbor, Michigan.
- POWELL, CHARLES A., 750 Harrison Avenue, Boston 18, Massachusetts.
- POWER, MAXWELL E., Department of Biology, Kenyon College, Gambier, Ohio.
- PRATT, IVAN, Department of Zoology, Oregon State College, Corvallis, Oregon.
- *PRICE, EMMETT W., Zoological Division, U. S. Bureau of Animal Industry, Beltsville, Maryland.
- PRICE, MANNING A., Department of Entomology, Texas A. and M. College, College Station, Texas.
- RAECKE, MARJORIE JEAN, Zoological Division, U. S. Bureau of Animal Industry, Beltsville, Maryland.
- RAMIREZ, J. J., Dean of Veterinary School, Caracas, Venezuela.
- RANKIN, JOHN S., JR., Department of Zoology, University of Connecticut, Storrs, Connecticut.
- RAPPAPORT, IRVING, 864 Cauldwell Avenue, New York, New York.
- RATCLIFFE, HERBERT L., Department of Pathology, University of Pennsylvania, Philadelphia, Pennsylvania.
- RAUSCH, ROBERT, Department of Veterinary Science, College of Agriculture, University of Wisconsin, Madison 6, Wisconsin.
- RAWSON, GEORGE W., Research Laboratories, Ciba Pharmaceutical Products Inc., Summit, New Jersey.
- READ, CLARK P., Department of Biology, Rice Institute, Houston, Texas.
- REARDON, LUCY, 805 Prince Street, Alexandria, Virginia.
- REDMOND, WILLIAM B., Box 534, Emory University, Atlanta, Georgia.
- REES, CHARLES W., National Institute of Health, 26th and East Streets, N.W., Washington, D. C.
- REESE, JOHN D., Department of Pathology and Bacteriology, University of Mississippi, School of Medicine, University, Mississippi.
- REIBER, ROBERT J., Department of Zoology, University of Georgia, Athens, Georgia.
- REID, MALCOLM W., Department of Biology, Monmouth College, Monmouth, Illinois.
- REIN, CHARLES ROBERT, 580 Fifth Avenue, New York 19, New York.
- *REYNOLDS, BRUCE D., Biology Department, University of Virginia, University Station, Charlottesville, Virginia.
- *RHODES, ROBERT C., Department of Biology, Emory University, University, Georgia.
- RICHARDSON, LAURENCE ROBERT, Department of Biology, Victoria University College, Wellington, W-1, New Zealand.
- RIEDEL, BERNARD B., School of Public Health, University of North Carolina, Chapel Hill, North Carolina.
- *RIETZ, JOHN H., Oglebay Hall, West Virginia University, Morgantown, West Virginia.
- *RILEY, WILLIAM A., Department of Zoology, University of Minnesota, Minneapolis, Minnesota.
- RISER, NATHAN W., Hopkins Marine Station, Pacific Grove, California.
- RITCHIE, LAWRENCE S., 406th General Medical Laboratory, APO 500, c/o Post Master, San Francisco, California.
- RITTERSON, ALBERT L., 307 Handy Street, New Brunswick, N. J.
- RODANICHE, ENID D., Box 466, Ancon, Canal Zone.
- ROSE, HARRY MELVIN, Department of Medicine, Columbia University, College of Physicians and Surgeons, New York 32, New York.
- ROSEBOOM, LLOYD E., Department of Parasitology, Johns Hopkins University, Baltimore, Maryland.
- ROSENAU, BARBARA J., School of Public Health, Chapel Hill, North Carolina.
- ROTHMAN, MAURICE M., 1727 Spruce Street, Philadelphia, Pennsylvania.

- ROUDABUSH, ROBERT L., Ward's Natural Science Establishment, 302 Goodman Street, Rochester, New York.
- ROZYCKI, ANTHONY T., 631 Stimson Street, Detroit, Michigan.
- RUBIN, GERARD, College of Veterinary Medicine, University of Illinois, Urbana, Illinois.
- RUSSELL, CATHERINE M., Department of Bacteriology and Parasitology, New York Medical College, 100th Street and Fifth Avenue, New York, New York.
- RUTHERFORD, ROBERT L., 635 W. Exposition Boulevard, Los Angeles, California.
- RYERSON, DWIGHT L., Department of Biology, Pomona College, Claremont, California.
- SACHS, IRVING BENJAMIN, Department of Zoology and Physiology, University of Illinois, Urbana, Illinois.
- SADUM, ELVIO H., 615 N. Wolfe Street, Baltimore 5, Maryland.
- SALSBUURY, JOHN G., Dr. Salsbury's Laboratories, Charles City, Iowa.
- SANDERS, MARGARET L., 609 College Avenue, Modesto, California.
- SANES, SAMUEL, Meyer Memorial and Children's Hospital, University of Buffalo Medical School, Buffalo, New York.
- SARLES, MERRITT P., 411 Montgomery Road, Laurel, Maryland.
- *SAUNDERS, G. LESLIE, University of Saskatchewan, Saskatoon, Canada.
- SAWITZ, WILLIAM G., Department of Bacteriology and Immunology, Jefferson Medical College, 1025 Walnut Street, Philadelphia 7, Pennsylvania.
- *SAWYER, WILBUR A., 3927 Idaho Avenue, N.W., Washington 8, D. C.
- SAXE, LEROY HALLOWELL, JR., 206 Bewley Road, Llanerch, Havertown, Pennsylvania.
- SCHIEFF, GEORGE J., Medical Research-Keneman Hall, Ohio State University, Columbus, Ohio.
- *SCHELL, MARGARET WOOD, University of Utah, Salt Lake City, Utah.
- SCHINAZI, LEWIS ARNOLD, Department of Zoology, University of California at Los Angeles, Los Angeles, California.
- SCHNEIDER, MORRIS D., 7914 Euclid Avenue, Chicago, Illinois.
- SCHUCK, BETTY RUTH, Department of Zoology, University of California, Berkeley, California.
- *SCHWARTZ, BENJAMIN, Zoological Division, U. S. Bureau of Animal Industry, Washington, D. C.
- *SCOTT, J. ALLEN, Department of Preventive Medicine, University of Texas Medical School, Galveston, Texas.
- *SCOTT, JOHN W., 1409 Garfield Street, Laramie, Wyoming.
- SEAMSTER, AARON, Department of Biology, University of Notre Dame, Notre Dame, Indiana.
- SEITNER, PHILIP G., Department of Biology, Purdue University, W. Lafayette, Indiana.
- SELF, JOHN TEAGUE, Department of Zoology, University of Oklahoma, Norman, Oklahoma.
- SHELANSKI, HERMAN A., Smyth Laboratories, 34th Street below Chestnut, Philadelphia, Pennsylvania.
- SHELDON, ALBERT J., Box #3, Norlina, North Carolina.
- *SHIELDS, RANDOLPH J., Library School of Medicine, Tsinan Shantung, China.
- SHORB, DOYS A., Zoological Division, U. S. Bureau of Animal Industry, Beltsville, Maryland.
- SHORT, ROBERT BROWN, Zoology Department, University of Michigan, Ann Arbor, Michigan.
- SHRAPNEL, BLISS C., United Fruit Company Hospital, Puerto Armuelles, Republic of Panama.
- SIMMS, BENNETT T., U. S. Bureau of Animal Industry, Washington, D. C.
- SIMPSON, MYRON L., Department of Biology, Gettysburg College, Gettysburg, Pennsylvania.
- SIMPSON, WILLIAM F., Department of Biology, The Catholic University of America, Washington, D. C.
- SMITH, BARNETT F., Spelman College, Atlanta, Georgia.
- SMITH, MELTON S., Chief of Medicine Service, 153rd Station Hospital, APO #75, % Post Master, San Francisco, California.
- *SMITH, SEPTIMA, University of Alabama, Box 1446, University, Alabama.
- SMITH, PHILIP EDWARD, 615 North Wolfe Street, Baltimore, Maryland.
- SMITH, VIVIAN S., Department of Parasitology, School of Hygiene and Public Health, Johns Hopkins University, Baltimore 5, Maryland.
- SOLANO GEISLER, SISTER FRANCIS, Nazareth College, Brighton Station, Rochester 10, New York.
- *SOMMERMEYER, VIOLA, 301 Medico-Dental Building, San Diego, California.
- SPIES, TOM DOUGLAS, Nutrition Clinic Hillman Hospital, Birmingham 3, Alabama.
- SPINDLER, LLOYD A., Zoological Division, U. S. Bureau of Animal Industry, Beltsville, Maryland.
- SPINGARN, CLIFFORD L., 898 Madison Avenue, New York 21, New York.
- SPRAGUE, VICTOR, Department of Zoology, Louisiana State University, Baton Rouge, Louisiana.
- STABLER, ROBERT M., Department of Biology, Colorado College, Colorado Springs, Colorado.
- *STAFFORD, ETHELBERT W., Agriculture and Mechanical College, State College, Mississippi.
- STAUBER, LESLIE A., Department of Zoology, Rutgers University, New Brunswick, New Jersey.
- *STEINER, GOTTHOLD, Division of Nematology, Bureau of Plant Industry, Washington, D. C.
- STEVENSON, ROBERT T., Department of Biology, University of Utah, Salt Lake City 1, Utah.

- *STEWART, M. A., 112 Agricultural Hall, University of California, Berkeley, California.
- *STOLL, NORMAN R., Rockefeller Institute for Medical Research, Princeton, New Jersey.
- STORCH, SIDNEY, Mt. Sinai Hospital, Fifth Avenue and 100th Street, New York, New York.
- STOVER, RICHARD D., Box 813, Iowa City, Iowa.
- STRANDTMANN, RUSSELL WILLIAM, Department of Public Health and Preventive Medicine, University of Texas Medical School, Galveston, Texas.
- *STUNKARD, HORACE W., Department of Biology, New York University, University Heights, New York 53, New York.
- SUMMERS, WILLIAM A., Medical College of Virginia, Richmond, Virginia.
- SWANSON, LEONARD E., Agricultural Experiment Station, Department of Animal Industry, University of Florida, Gainesville, Florida.
- SWARTZWELDER, JOHN CLYDE, Department of Public Health, School of Medicine, Louisiana State University, 1542 Tulane Avenue, New Orleans 13, Louisiana.
- SWEZY, WILLIAM W., Department of Zoology, Grove City College, Grove City, Pennsylvania.
- TAKOS, MICHAEL JAMES, Department of Biology, Emory University, Atlanta, Georgia.
- *TALIAFERRO, WILLIAM H., Department of Bacteriology and Parasitology, University of Chicago, Chicago, Illinois.
- TAYLOR, LELAND HART, Department of Zoology, West Virginia University, Morgantown, West Virginia.
- TENBROECK, CARL, Rockefeller Institute for Medical Research, Princeton, New Jersey.
- TERZIAN, LEVON A., Naval Medical Research Institute, Bethesda 14, Maryland.
- TETLEY, JOHN H., Massey Agricultural College, Palmerston North, New Zealand.
- *THOMAS, LYLE J., 103 Vivarium Building, Champaign, Illinois.
- *THOMPSON, PAUL E., Research Division, Parke Davis and Company, Detroit, Michigan.
- THRELKELD, WILLIAM L., Virginia Agricultural Experiment Station, Blacksburg, Virginia.
- THURMAN, DEED C., JR., P. O. Box 210, Florida State Board of Health, Jacksonville, Florida.
- TINER, JACK D., Department of Veterinary Science, University of Wisconsin, Madison, Wisconsin.
- TODD, ARLIE C., University of Tennessee, Agricultural Experiment Station, Knoxville 16, Tennessee.
- TOWER, NANCY M., 35 North Main Street, Cohasset, Massachusetts.
- TOWNSEND, ELSIE W., Department of Biology, Wayne University, Detroit, Michigan.
- TRAGER, WILLIAM, Rockefeller Institute for Medical Research, Princeton, New Jersey.
- TRAUB, ROBERT, Entomology Section, Division of Parasitology, Army Medical School, Washington, D. C.
- TRAVIS, BERNARD, Box 3391, Orlando, Florida.
- TREMBLEY, HELEN LOUISE, Malaria Laboratory, National Institute of Health, Bethesda, Maryland.
- TROWBRIDGE, MINNIE S., Zoological Division, Bureau of Animal Industry, Beltsville, Maryland.
- TSUCHIYA, HIROMU, Department of Bacteriology and Hygiene, Washington University, Saint Louis, Missouri.
- TUCKER, HAROLD, Johns Hopkins Hospital, Baltimore 5, Maryland.
- TULLOCH, GEORGE SHERLOCK, Department of Biology, Brooklyn College, Brooklyn, New York.
- TURK, RICHARD DUNCAN, Department of Veterinary Parasitology, Texas Agricultural and Mechanical College, College Station, Texas.
- *TYZZER, ERNEST E., Department of Comparative Pathology, Harvard Medical School, Boston, Massachusetts.
- *VAN CLEAVE, HARLEY J., Department of Zoology, University of Illinois, Urbana, Illinois.
- VAN DER WOUDE, ANNE, Department of Zoology, University of Michigan, Ann Arbor, Michigan.
- VARGAS, LUIS, National Faculty of Medicine, Calle de la Escuela, Medico-Militar No. 20, Mexico City, Mexico.
- VAUGHN, CHARLES M., Department of Zoology, Miami University, Oxford, Ohio.
- VENARD, CARL E., Department of Zoology and Entomology, Ohio State University, Columbus, Ohio.
- VOGE, MARIETTA, Department of Zoology, University of California, Berkeley, California.
- VOGEL, HANS, Institut für Schiffs-und Tropenkrankheiten, Bernhard-Nochtstrasse 74, Hamburg 4, Germany.
- VOLK, JOSEPH J., 5931 West Michigan Street, Milwaukee 13, Wisconsin.
- VON BRAND, THEODOR, Division of Zoology, National Institute of Health, Bethesda 14, Maryland.
- WAGNER, EDWARD D., Biology Department, Atlantic Union College, S. Lancaster, Massachusetts.
- WALETZLSY, EMANUEL, Stamford Research Laboratories, American Cyanamid Company, Stamford, Connecticut.
- WALKER, J. HENRY, Department of Biology, University of Alabama, University, Alabama.
- WALLACE, FRANKLIN G., 3349 University Avenue, S.E., Minneapolis 14, Minnesota.

- WALLACE, HAROLD E., Michigan State Normal College, Ypsilanti, Michigan.
- *WALTON, ARTHUR C., Department of Biology, Knox College, Galesburg, Illinois.
- WANG, JOAN SHU HSU, Department of Zoology, Louisiana State University, Baton Rouge, Louisiana.
- WANTLAND, WAYNE W., Department of Biology and Health Science, Illinois Wesleyan University, Bloomington, Illinois.
- WARD, HELEN L., College of Liberal Arts, University of Tennessee, Knoxville, Tennessee.
- WARD, JAMES W., Box 183, University, Mississippi.
- WARE, ROBERT E., Clemson Agricultural and Mechanical College, Clemson, South Carolina.
- *WARREN, ANDREW J., The Rockefeller Foundation, 49 West 49th Street, New York City, New York.
- WARREN, HERBERT S., Department of Anatomy, Hahneman Medical College, 235 N. 15th Street, Philadelphia 2, Pennsylvania.
- WEATHERS, CURTIS L., Department of Biology, Long Island University, 300 Pearl Street, Brooklyn, New York.
- WEBSTER, JACKSON DAN, Department of Biology, Rice Institute, Houston, Texas.
- *WEHR, EVERETT EIMER, Zoological Division, U. S. Bureau of Animal Industry, Beltsville, Maryland.
- WEINMAN, DAVID, 33 South Russell Street, Boston 14, Massachusetts.
- WEINSTEIN, PAUL P., Department of Parasitology, Johns Hopkins University, Baltimore 5, Maryland.
- WELLER, THOMAS H., 8 Netherlands Road, Brookline, Massachusetts.
- *WENRICH, DAVID H., Zoology Department, University of Pennsylvania, Philadelphia, Pennsylvania.
- WERBY, HELEN J., Department of Biology, Seattle College, Seattle 22, Washington.
- WEST, EVELYN D., State Department of Health, P. O. Box 1139, Hartford, Connecticut.
- WEARTON, DENNIS R. A., Lankenau Research Institute, Girard and Corinthian Streets, Philadelphia, Pennsylvania.
- WHARTON, GEORGE W., Zoology Department, Duke University, Durham, North Carolina.
- WHATLEY, EDWARD C., Department of Biology, Northeast Junior College, Monroe, Louisiana.
- WHITENER, PAUL D., Cramerton, North Carolina.
- WHITLOCK, JOHN H., State Veterinary College, Cornell University, Ithaca, New York.
- *WHITMORE, EUGENE R., 2139 Wyoming Avenue, N.W., Washington, D. C.
- WHITT, ALLIE LOUIS, JR., P. O. Box 1771, University Station, Lexington, Kentucky.
- WICHTERMAN, RALPH, Biology Department, Temple University, Philadelphia, Pennsylvania.
- WILHELM, RAYMOND W., 50 Haven Avenue, New York 32, New York.
- WILEY, CHARLES H., Department of Biology, New York University, University Heights, New York 52, New York.
- WILLIAM, THOMAS H., Deerlodge Hospital, Winnipeg, Canada.
- WILLIAMS, ROGER WRIGHT, De Lamar Institute of Public Health, Columbia University, New York, New York.
- WILLIAMS, THEODORE S., P. O. Box 893, Tuskegee Institute, Alabama.
- WILMOTH, JAMES H., Department of Biology, Brooklyn College, Brooklyn, New York.
- WOKE, PAUL A., U. S. Public Health Service, P. O. Box 547, Savannah, Georgia.
- WOLFSON, FRUMA, Department of Parasitology, Johns Hopkins University, Baltimore 9, Maryland.
- WOMERSLEY, H., South Australian Museum, Adelaide, South Australia.
- WOOD, SHERWIN F., 1015 North Alexandria, Hollywood 27, California.
- WOODHEAD, A. E., Department of Zoology, University of Michigan, Ann Arbor, Michigan.
- WORTH, C. BROOKE, 107 Parker Street, Tampa, Florida.
- WRIGHT, WILLARD H., National Institute of Health, Tropical Disease Laboratory, U. S. Public Health Service, Bethesda, Maryland.
- WYBORNEY, EUGENE H., 219 6th Avenue, Kirkland, Washington.
- YOLLES, STANLEY F., 189 E. 18th Street, Brooklyn, New York.
- YOLLES, TAMARATH KNIGIN, Department of Preventive Medicine, New York University, 477 First Avenue, New York 16, New York.
- YOUNG, BENJAMIN P., Department of Zoology, Cornell University, Ithaca, New York.
- YOUNG, MARTIN D., U. S. Public Health Service, Box 1344, Columbia, South Carolina.
- YOUNG, VIOLA MAE, Research Laboratory, Mount Sinai Hospital, Chicago 8, Illinois.
- YUTUC, LOPE M., Department of Veterinary Medicine, College of Veterinary Science, University of Philippines, Manila, Philippine Islands.
- ZELIFF, C. COURSON, Department of Zoology, Pennsylvania State College, State College, Pennsylvania.
- *ZETEK, JAMES, Canal Zone Biological Area, Drawer C, Balboa, Canal Zone.
- ZUCKER, ISADORE, 112 Shanley Avenue, Newark 8, New Jersey.

AMERICAN SOCIETY OF PARASITOLOGISTS

ANNOUNCEMENTS

Horace W. Stunkard, who has served as Chairman of the Editorial Committee of the Journal of Parasitology for the past four years, will have sabbatical leave from New York University from February to September, 1948. Consequently, W. W. Cort will serve as Chairman of the Editorial Committee and managing editor of the Journal of Parasitology during 1948 (volume 34). Manuscripts and correspondence concerning editorial affairs may be addressed to him: Professor W. W. Cort, Department of Parasitology, Johns Hopkins University, Baltimore 5, Maryland.

Mailing dates of volume 32, Journal of Parasitology

February issue		April 22, 1946
April issue		May 22, 1946
June issue		July 24, 1946
August issue		September 5, 1946
October issue		October 29, 1946
December issue	Section 1	February 26, 1947
	Section 2	December 17, 1946